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(54) Title: TRANSPORTERS AND ION CHANNELS

(57) Abstract: The invention provides human transporters and ion channels (TRICH) and polynucleotides which identify and encode TRICH. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of TRICH.

TRANSPORTERS AND ION CHANNELS

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of transporters and ion channels and to the use of these sequences in the diagnosis, treatment, and prevention of transport, neurological, muscle, and immunological disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

BACKGROUND OF THE INVENTION

Eukaryotic cells are surrounded and subdivided into functionally distinct organelles by hydrophobic lipid bilayer membranes which are highly impermeable to most polar molecules. Cells and organelles require transport proteins to import and export essential nutrients and metal ions including K^+ , NH_4^+ , P_i , SO_4^{2-} , sugars, and vitamins, as well as various metabolic waste products. Transport proteins also play roles in antibiotic resistance, toxin secretion, ion balance, synaptic neurotransmission, kidney function, intestinal absorption, tumor growth, and other diverse cell functions (Griffith, J. and C. Sansom (1998) The Transporter Facts Book, Academic Press, San Diego CA, pp. 3-29). Transport can occur by a passive concentration-dependent mechanism, or can be linked to an energy source such as ATP hydrolysis or an ion gradient. Proteins that function in transport include carrier proteins, which bind to a specific solute and undergo a conformational change that translocates the bound solute across the membrane, and channel proteins, which form hydrophilic pores that allow specific solutes to diffuse through the membrane down an electrochemical solute gradient.

Carrier proteins which transport a single solute from one side of the membrane to the other are called uniporters. In contrast, coupled transporters link the transfer of one solute with simultaneous or sequential transfer of a second solute, either in the same direction (symport) or in the opposite direction (antiport). For example, intestinal and kidney epithelium contains a variety of symporter systems driven by the sodium gradient that exists across the plasma membrane. Sodium moves into the cell down its electrochemical gradient and brings the solute into the cell with it. The sodium gradient that provides the driving force for solute uptake is maintained by the ubiquitous Na^+/K^+ ATPase system. Sodium-coupled transporters include the mammalian glucose transporter (SGLT1), iodide transporter (NIS), and multivitamin transporter (SMVT). All three transporters have twelve putative transmembrane segments, extracellular glycosylation sites, and cytoplasmically-oriented N- and C-termini. NIS plays a crucial role in the evaluation, diagnosis, and treatment of various thyroid pathologies because it is the molecular basis for radioiodide thyroid-imaging techniques and for specific targeting of radioisotopes to the thyroid gland (Levy, O. et al. (1997) Proc.

Natl. Acad. Sci. USA 94:5568-5573). SMVT is expressed in the intestinal mucosa, kidney, and placenta, and is implicated in the transport of the water-soluble vitamins, e.g., biotin and pantothenate (Prasad, P.D. et al. (1998) J. Biol. Chem. 273:7501-7506).

One of the largest families of transporters is the major facilitator superfamily (MFS), also called the uniporter-symporter-antiporter family. MFS transporters are single polypeptide carriers that transport small solutes in response to ion gradients. Members of the MFS are found in all classes of living organisms, and include transporters for sugars, oligosaccharides, phosphates, nitrates, nucleosides, monocarboxylates, and drugs. MFS transporters found in eukaryotes all have a structure comprising 12 transmembrane segments (Pao, S.S. et al. (1998) Microbiol. Molec. Biol. Rev. 62:1-34). The largest family of MFS transporters is the sugar transporter family, which includes the seven glucose transporters (GLUT1-GLUT7) found in humans that are required for the transport of glucose and other hexose sugars. These glucose transport proteins have unique tissue distributions and physiological functions. GLUT1 provides many cell types with their basal glucose requirements and transports glucose across epithelial and endothelial barrier tissues; GLUT2 facilitates glucose uptake or efflux from the liver; GLUT3 regulates glucose supply to neurons; GLUT4 is responsible for insulin-regulated glucose disposal; and GLUT5 regulates fructose uptake into skeletal muscle. Defects in glucose transporters are involved in a recently identified neurological syndrome causing infantile seizures and developmental delay, as well as glycogen storage disease, Fanconi-Bickel syndrome, and non-insulin-dependent diabetes mellitus (Mueckler, M. (1994) Eur. J. Biochem. 219:713-725; Longo, N. and L.J. Elsas (1998) Adv. Pediatr. 45:293-313).

Monocarboxylate anion transporters are proton-coupled symporters with a broad substrate specificity that includes L-lactate, pyruvate, and the ketone bodies acetate, acetoacetate, and beta-hydroxybutyrate. At least seven isoforms have been identified to date. The isoforms are predicted to have twelve transmembrane (TM) helical domains with a large intracellular loop between TM6 and TM7, and play a critical role in maintaining intracellular pH by removing the protons that are produced stoichiometrically with lactate during glycolysis. The best characterized H⁺-monocarboxylate transporter is that of the erythrocyte membrane, which transports L-lactate and a wide range of other aliphatic monocarboxylates. Other cells possess H⁺-linked monocarboxylate transporters with differing substrate and inhibitor selectivities. In particular, cardiac muscle and tumor cells have transporters that differ in their K_m values for certain substrates, including stereoselectivity for L- over D-lactate, and in their sensitivity to inhibitors. There are Na⁺-monocarboxylate cotransporters on the luminal surface of intestinal and kidney epithelia, which allow the uptake of lactate, pyruvate, and ketone bodies in these tissues. In addition, there are specific and selective transporters for organic cations and organic anions in organs including the kidney, intestine and liver. Organic anion transporters are selective for

hydrophobic, charged molecules with electron-attracting side groups. Organic cation transporters, such as the ammonium transporter, mediate the secretion of a variety of drugs and endogenous metabolites, and contribute to the maintenance of intercellular pH (Poole, R.C. and A.P. Halestrap (1993) *Am. J. Physiol.* 264:C761-C782; Price, N.T. et al. (1998) *Biochem. J.* 329:321-328; and Martinelle, K. and I. Haggstrom (1993) *J. Biotechnol.* 30:339-350).

ATP-binding cassette (ABC) transporters are members of a superfamily of membrane proteins that transport substances ranging from small molecules such as ions, sugars, amino acids, peptides, and phospholipids, to lipopeptides, large proteins, and complex hydrophobic drugs. ABC transporters consist of four modules: two nucleotide-binding domains (NBD), which hydrolyze ATP to supply the energy required for transport, and two membrane-spanning domains (MSD), each containing six putative transmembrane segments. These four modules may be encoded by a single gene, as is the case for the cystic fibrosis transmembrane regulator (CFTR), or by separate genes. When encoded by separate genes, each gene product contains a single NBD and MSD. These "half-molecules" form homo- and heterodimers, such as Tap1 and Tap2, the endoplasmic reticulum-based major histocompatibility (MHC) peptide transport system. Several genetic diseases are attributed to defects in ABC transporters, such as the following diseases and their corresponding proteins: cystic fibrosis (CFTR, an ion channel), adrenoleukodystrophy (adrenoleukodystrophy protein, ALDP), Zellweger syndrome (peroxisomal membrane protein-70, PMP70), and hyperinsulinemic hypoglycemia (sulfonylurea receptor, SUR). Overexpression of the multidrug resistance (MDR) protein, another ABC transporter, in human cancer cells makes the cells resistant to a variety of cytotoxic drugs used in chemotherapy (Taglicht, D. and S. Michaelis (1998) *Meth. Enzymol.* 292:130-162).

A number of metal ions such as iron, zinc, copper, cobalt, manganese, molybdenum, selenium, nickel, and chromium are important as cofactors for a number of enzymes. For example, copper is involved in hemoglobin synthesis, connective tissue metabolism, and bone development, by acting as a cofactor in oxidoreductases such as superoxide dismutase, ferroxidase (ceruloplasmin), and lysyl oxidase. Copper and other metal ions must be provided in the diet, and are absorbed by transporters in the gastrointestinal tract. Plasma proteins transport the metal ions to the liver and other target organs, where specific transporters move the ions into cells and cellular organelles as needed. Imbalances in metal ion metabolism have been associated with a number of disease states (Danks, D.M. (1986) *J. Med. Genet.* 23:99-106).

Transport of fatty acids across the plasma membrane can occur by diffusion, a high capacity, low affinity process. However, under normal physiological conditions a significant fraction of fatty acid transport appears to occur via a high affinity, low capacity protein-mediated transport process. Fatty acid transport protein (FATP), an integral membrane protein with four transmembrane segments,

is expressed in tissues exhibiting high levels of plasma membrane fatty acid flux, such as muscle, heart, and adipose. Expression of FATP is upregulated in 3T3-L1 cells during adipose conversion, and expression in COS7 fibroblasts elevates uptake of long-chain fatty acids (Hui, T.Y. et al. (1998) J. Biol. Chem. 273:27420-27429).

5 Mitochondrial carrier proteins are transmembrane-spanning proteins which transport ions and charged metabolites between the cytosol and the mitochondrial matrix. Examples include the ADP, ATP carrier protein; the 2-oxoglutarate/malate carrier; the phosphate carrier protein; the pyruvate carrier; the dicarboxylate carrier which transports malate, succinate, fumarate, and phosphate; the tricarboxylate carrier which transports citrate and malate; and the Grave's disease carrier protein, a
10 protein recognized by IgG in patients with active Grave's disease, an autoimmune disorder resulting in hyperthyroidism. Proteins in this family consist of three tandem repeats of an approximately 100 amino acid domain, each of which contains two transmembrane regions (Stryer, L. (1995) Biochemistry, W.H. Freeman and Company, New York NY, p. 551; PROSITE PDOC00189 Mitochondrial energy transfer proteins signature; Online Mendelian Inheritance in Man (OMIM) *275000 Graves Disease).

15 This class of transporters also includes the mitochondrial uncoupling proteins, which create proton leaks across the inner mitochondrial membrane, thus uncoupling oxidative phosphorylation from ATP synthesis. The result is energy dissipation in the form of heat. Mitochondrial uncoupling proteins have been implicated as modulators of thermoregulation and metabolic rate, and have been proposed as potential targets for drugs against metabolic diseases such as obesity (Ricquier, D. et al. (1999) J. Int.
20 Med. 245:637-642).

Ion Channels

The electrical potential of a cell is generated and maintained by controlling the movement of ions across the plasma membrane. The movement of ions requires ion channels, which form ion-selective pores within the membrane. There are two basic types of ion channels, ion transporters and
25 gated ion channels. Ion transporters utilize the energy obtained from ATP hydrolysis to actively transport an ion against the ion's concentration gradient. Gated ion channels allow passive flow of an ion down the ion's electrochemical gradient under restricted conditions. Together, these types of ion channels generate, maintain, and utilize an electrochemical gradient that is used in 1) electrical impulse conduction down the axon of a nerve cell, 2) transport of molecules into cells against concentration
30 gradients, 3) initiation of muscle contraction, and 4) endocrine cell secretion.

Ion Transporters

Ion transporters generate and maintain the resting electrical potential of a cell. Utilizing the energy derived from ATP hydrolysis, they transport ions against the ion's concentration gradient. These transmembrane ATPases are divided into three families. The phosphorylated (P) class ion

transporters, including Na^+ - K^+ ATPase, Ca^{2+} -ATPase, and H^+ -ATPase, are activated by a phosphorylation event. P-class ion transporters are responsible for maintaining resting potential distributions such that cytosolic concentrations of Na^+ and Ca^{2+} are low and cytosolic concentration of K^+ is high. The vacuolar (V) class of ion transporters includes H^+ pumps on intracellular organelles, such as lysosomes and Golgi. V-class ion transporters are responsible for generating the low pH within the lumen of these organelles that is required for function. The coupling factor (F) class consists of H^+ pumps in the mitochondria. F-class ion transporters utilize a proton gradient to generate ATP from ADP and inorganic phosphate (P_i).

The P-ATPases are hexamers of a 100 kD subunit with ten transmembrane domains and several large cytoplasmic regions that may play a role in ion binding (Scarborough, G.A. (1999) *Curr. Opin. Cell Biol.* 11:517-522). The V-ATPases are composed of two functional domains: the V_1 domain, a peripheral complex responsible for ATP hydrolysis; and the V_0 domain, an integral complex responsible for proton translocation across the membrane. The F-ATPases are structurally and evolutionarily related to the V-ATPases. The F-ATPase F_0 domain contains 12 copies of the c subunit, a highly hydrophobic protein composed of two transmembrane domains and containing a single buried carboxyl group in TM2 that is essential for proton transport. The V-ATPase V_0 domain contains three types of homologous c subunits with four or five transmembrane domains and the essential carboxyl group in TM4 or TM3. Both types of complex also contain a single a subunit that may be involved in regulating the pH dependence of activity (Forgac, M. (1999) *J. Biol. Chem.* 274:12951-12954).

The resting potential of the cell is utilized in many processes involving carrier proteins and gated ion channels. Carrier proteins utilize the resting potential to transport molecules into and out of the cell. Amino acid and glucose transport into many cells is linked to sodium ion co-transport (symport) so that the movement of Na^+ down an electrochemical gradient drives transport of the other molecule up a concentration gradient. Similarly, cardiac muscle links transfer of Ca^{2+} out of the cell with transport of Na^+ into the cell (antiport).

Gated Ion Channels

Gated ion channels control ion flow by regulating the opening and closing of pores. The ability to control ion flux through various gating mechanisms allows ion channels to mediate such diverse signaling and homeostatic functions as neuronal and endocrine signaling, muscle contraction, fertilization, and regulation of ion and pH balance. Gated ion channels are categorized according to the manner of regulating the gating function. Mechanically-gated channels open their pores in response to mechanical stress; voltage-gated channels (e.g., Na^+ , K^+ , Ca^{2+} , and Cl^- channels) open their pores in response to changes in membrane potential; and ligand-gated channels (e.g., acetylcholine-, serotonin-, and glutamate-gated cation channels, and GABA- and glycine-gated

chloride channels) open their pores in the presence of a specific ion, nucleotide, or neurotransmitter. The gating properties of a particular ion channel (i.e., its threshold for and duration of opening and closing) are sometimes modulated by association with auxiliary channel proteins and/or post translational modifications, such as phosphorylation.

5 Mechanically-gated or mechanosensitive ion channels act as transducers for the senses of touch, hearing, and balance, and also play important roles in cell volume regulation, smooth muscle contraction, and cardiac rhythm generation. A stretch-inactivated channel (SIC) was recently cloned from rat kidney. The SIC channel belongs to a group of channels which are activated by pressure or stress on the cell membrane and conduct both Ca^{2+} and Na^{+} (Suzuki, M. et al. (1999) J. Biol. Chem.
10 274:6330-6335).

 The pore-forming subunits of the voltage-gated cation channels form a superfamily of ion channel proteins. The characteristic domain of these channel proteins comprises six transmembrane domains (S1-S6), a pore-forming region (P) located between S5 and S6, and intracellular amino and carboxy termini. In the Na^{+} and Ca^{2+} subfamilies, this domain is repeated four times, while in the K^{+}
15 channel subfamily, each channel is formed from a tetramer of either identical or dissimilar subunits. The P region contains information specifying the ion selectivity for the channel. In the case of K^{+} channels, a GYG tripeptide is involved in this selectivity (Ishii, T.M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11651-11656).

 Voltage-gated Na^{+} and K^{+} channels are necessary for the function of electrically excitable cells,
20 such as nerve and muscle cells. Action potentials, which lead to neurotransmitter release and muscle contraction, arise from large, transient changes in the permeability of the membrane to Na^{+} and K^{+} ions. Depolarization of the membrane beyond the threshold level opens voltage-gated Na^{+} channels. Sodium ions flow into the cell, further depolarizing the membrane and opening more voltage-gated Na^{+} channels, which propagates the depolarization down the length of the cell. Depolarization also opens
25 voltage-gated potassium channels. Consequently, potassium ions flow outward, which leads to repolarization of the membrane. Voltage-gated channels utilize charged residues in the fourth transmembrane segment (S4) to sense voltage change. The open state lasts only about 1 millisecond, at which time the channel spontaneously converts into an inactive state that cannot be opened irrespective of the membrane potential. Inactivation is mediated by the channel's N-terminus, which acts as a plug
30 that closes the pore. The transition from an inactive to a closed state requires a return to resting potential.

 Voltage-gated Na^{+} channels are heterotrimeric complexes composed of a 260 kDa pore-forming α subunit that associates with two smaller auxiliary subunits, $\beta 1$ and $\beta 2$. The $\beta 2$ subunit is a integral membrane glycoprotein that contains an extracellular Ig domain, and its association with α and $\beta 1$

subunits correlates with increased functional expression of the channel, a change in its gating properties, as well as an increase in whole cell capacitance due to an increase in membrane surface area (Isom, L.L. et al. (1995) *Cell* 83:433-442).

Non voltage-gated Na^+ channels include the members of the amiloride-sensitive Na^+ channel/degenerin (NaC/DEG) family. Channel subunits of this family are thought to consist of two transmembrane domains flanking a long extracellular loop, with the amino and carboxyl termini located within the cell. The NaC/DEG family includes the epithelial Na^+ channel (ENaC) involved in Na^+ reabsorption in epithelia including the airway, distal colon, cortical collecting duct of the kidney, and exocrine duct glands. Mutations in ENaC result in pseudohypoaldosteronism type 1 and Liddle's syndrome (pseudohyperaldosteronism). The NaC/DEG family also includes the recently characterized H^+ -gated cation channels or acid-sensing ion channels (ASIC). ASIC subunits are expressed in the brain and form heteromultimeric Na^+ -permeable channels. These channels require acid pH fluctuations for activation. ASIC subunits show homology to the degenerins, a family of mechanically-gated channels originally isolated from *C. elegans*. Mutations in the degenerins cause neurodegeneration. ASIC subunits may also have a role in neuronal function, or in pain perception, since tissue acidosis causes pain (Waldmann, R. and M. Lazdunski (1998) *Curr. Opin. Neurobiol.* 8:418-424; Eglen, R.M. et al. (1999) *Trends Pharmacol. Sci.* 20:337-342).

K^+ channels are located in all cell types, and may be regulated by voltage, ATP concentration, or second messengers such as Ca^{2+} and cAMP. In non-excitabile tissue, K^+ channels are involved in protein synthesis, control of endocrine secretions, and the maintenance of osmotic equilibrium across membranes. In neurons and other excitable cells, in addition to regulating action potentials and repolarizing membranes, K^+ channels are responsible for setting resting membrane potential. The cytosol contains non-diffusible anions and, to balance this net negative charge, the cell contains a Na^+ - K^+ pump and ion channels that provide the redistribution of Na^+ , K^+ , and Cl^- . The pump actively transports Na^+ out of the cell and K^+ into the cell in a 3:2 ratio. Ion channels in the plasma membrane allow K^+ and Cl^- to flow by passive diffusion. Because of the high negative charge within the cytosol, Cl^- flows out of the cell. The flow of K^+ is balanced by an electromotive force pulling K^+ into the cell, and a K^+ concentration gradient pushing K^+ out of the cell. Thus, the resting membrane potential is primarily regulated by K^+ flow (Salkoff, L. and T. Jegla (1995) *Neuron* 15:489-492).

Potassium channel subunits of the Shaker-like superfamily all have the characteristic six transmembrane/1 pore domain structure. Four subunits combine as homo- or heterotetramers to form functional K channels. These pore-forming subunits also associate with various cytoplasmic β subunits that alter channel inactivation kinetics. The Shaker-like channel family includes the voltage-gated K^+ channels as well as the delayed rectifier type channels such as the human ether-a-go-go

related gene (HERG) associated with long QT, a cardiac dysrhythmia syndrome (Curran, M.E. (1998) *Curr. Opin. Biotechnol.* 9:565-572; Kaczarowski, G.J. and M.L. Garcia (1999) *Curr. Opin. Chem. Biol.* 3:448-458).

A second superfamily of K⁺ channels is composed of the inward rectifying channels (Kir).
5 Kir channels have the property of preferentially conducting K⁺ currents in the inward direction. These proteins consist of a single potassium selective pore domain and two transmembrane domains, which correspond to the fifth and sixth transmembrane domains of voltage-gated K⁺ channels. Kir subunits also associate as tetramers. The Kir family includes ROMK1, mutations in which lead to Bartter syndrome, a renal tubular disorder. Kir channels are also involved in regulation of cardiac pacemaker
10 activity, seizures and epilepsy, and insulin regulation (Doupnik, C.A. et al. (1995) *Curr. Opin. Neurobiol.* 5:268-277; Curran, *supra*).

The recently recognized TWIK K⁺ channel family includes the mammalian TWIK-1, TREK-1 and TASK proteins. Members of this family possess an overall structure with four transmembrane domains and two P domains. These proteins are probably involved in controlling the resting potential
15 in a large set of cell types (Duprat, F. et al. (1997) *EMBO J* 16:5464-5471).

The voltage-gated Ca²⁺ channels have been classified into several subtypes based upon their electrophysiological and pharmacological characteristics. L-type Ca²⁺ channels are predominantly expressed in heart and skeletal muscle where they play an essential role in excitation-contraction coupling. T-type channels are important for cardiac pacemaker activity, while N-type and P/Q-type
20 channels are involved in the control of neurotransmitter release in the central and peripheral nervous system. The L-type and N-type voltage-gated Ca²⁺ channels have been purified and, though their functions differ dramatically, they have similar subunit compositions. The channels are composed of three subunits. The α_1 subunit forms the membrane pore and voltage sensor, while the $\alpha_2\delta$ and β subunits modulate the voltage-dependence, gating properties, and the current amplitude of the channel.
25 These subunits are encoded by at least six α_1 , one $\alpha_2\delta$, and four β genes. A fourth subunit, γ , has been identified in skeletal muscle (Walker, D. et al. (1998) *J. Biol. Chem.* 273:2361-2367; McCleskey, E.W. (1994) *Curr. Opin. Neurobiol.* 4:304-312).

Chloride channels are necessary in endocrine secretion and in regulation of cytosolic and organelle pH. In secretory epithelial cells, Cl⁻ enters the cell across a basolateral membrane through an
30 Na⁺, K⁺/Cl⁻ cotransporter, accumulating in the cell above its electrochemical equilibrium concentration. Secretion of Cl⁻ from the apical surface, in response to hormonal stimulation, leads to flow of Na⁺ and water into the secretory lumen. The cystic fibrosis transmembrane conductance regulator (CFTR) is a chloride channel encoded by the gene for cystic fibrosis, a common fatal genetic disorder in humans. CFTR is a member of the ABC transporter family, and is composed of two domains each consisting of

six transmembrane domains followed by a nucleotide-binding site. Loss of CFTR function decreases transepithelial water secretion and, as a result, the layers of mucus that coat the respiratory tree, pancreatic ducts, and intestine are dehydrated and difficult to clear. The resulting blockage of these sites leads to pancreatic insufficiency, "meconium ileus", and devastating "chronic obstructive pulmonary disease" (Al-Awqati, Q. et al. (1992) J. Exp. Biol. 172:245-266).

The voltage-gated chloride channels (CLC) are characterized by 10-12 transmembrane domains, as well as two small globular domains known as CBS domains. The CLC subunits probably function as homotetramers. CLC proteins are involved in regulation of cell volume, membrane potential stabilization, signal transduction, and transepithelial transport. Mutations in CLC-1, expressed predominantly in skeletal muscle, are responsible for autosomal recessive generalized myotonia and autosomal dominant myotonia congenita, while mutations in the kidney channel CLC-5 lead to kidney stones (Jentsch, T.J. (1996) Curr. Opin. Neurobiol. 6:303-310).

Ligand-gated channels open their pores when an extracellular or intracellular mediator binds to the channel. Neurotransmitter-gated channels are channels that open when a neurotransmitter binds to their extracellular domain. These channels exist in the postsynaptic membrane of nerve or muscle cells. There are two types of neurotransmitter-gated channels. Sodium channels open in response to excitatory neurotransmitters, such as acetylcholine, glutamate, and serotonin. This opening causes an influx of Na^+ and produces the initial localized depolarization that activates the voltage-gated channels and starts the action potential. Chloride channels open in response to inhibitory neurotransmitters, such as γ -aminobutyric acid (GABA) and glycine, leading to hyperpolarization of the membrane and the subsequent generation of an action potential. Neurotransmitter-gated ion channels have four transmembrane domains and probably function as pentamers (Jentsch, supra). Amino acids in the second transmembrane domain appear to be important in determining channel permeation and selectivity (Sather, W.A. et al. (1994) Curr. Opin. Neurobiol. 4:313-323).

Ligand-gated channels can be regulated by intracellular second messengers. For example, calcium-activated K^+ channels are gated by internal calcium ions. In nerve cells, an influx of calcium during depolarization opens K^+ channels to modulate the magnitude of the action potential (Ishi et al., supra). The large conductance (BK) channel has been purified from brain and its subunit composition determined. The α subunit of the BK channel has seven rather than six transmembrane domains in contrast to voltage-gated K^+ channels. The extra transmembrane domain is located at the subunit N-terminus. A 28-amino-acid stretch in the C-terminal region of the subunit (the "calcium bowl" region) contains many negatively charged residues and is thought to be the region responsible for calcium binding. The β subunit consists of two transmembrane domains connected by a glycosylated extracellular loop, with intracellular N- and C-termini (Kaczorowski, supra; Vergara, C. et al. (1998)

Curr. Opin. Neurobiol. 8:321-329).

Cyclic nucleotide-gated (CNG) channels are gated by cytosolic cyclic nucleotides. The best examples of these are the cAMP-gated Na⁺ channels involved in olfaction and the cGMP-gated cation channels involved in vision. Both systems involve ligand-mediated activation of a G-protein coupled receptor which then alters the level of cyclic nucleotide within the cell. CNG channels also represent a major pathway for Ca²⁺ entry into neurons, and play roles in neuronal development and plasticity. CNG channels are tetramers containing at least two types of subunits, an α subunit which can form functional homomeric channels, and a β subunit, which modulates the channel properties. All CNG subunits have six transmembrane domains and a pore forming region between the fifth and sixth transmembrane domains, similar to voltage-gated K⁺ channels. A large C-terminal domain contains a cyclic nucleotide binding domain, while the N-terminal domain confers variation among channel subtypes (Zufall, F. et al. (1997) Curr. Opin. Neurobiol. 7:404-412).

The activity of other types of ion channel proteins may also be modulated by a variety of intracellular signalling proteins. Many channels have sites for phosphorylation by one or more protein kinases including protein kinase A, protein kinase C, tyrosine kinase, and casein kinase II, all of which regulate ion channel activity in cells. Kir channels are activated by the binding of the G $\beta\gamma$ subunits of heterotrimeric G-proteins (Reimann, F. and F.M. Ashcroft (1999) Curr. Opin. Cell. Biol. 11:503-508). Other proteins are involved in the localization of ion channels to specific sites in the cell membrane. Such proteins include the PDZ domain proteins known as MAGUKs (membrane-associated guanylate kinases) which regulate the clustering of ion channels at neuronal synapses (Craven, S.E. and D.S. Bredt (1998) Cell 93:495-498).

Disease Correlation

The etiology of numerous human diseases and disorders can be attributed to defects in the transport of molecules across membranes. Defects in the trafficking of membrane-bound transporters and ion channels are associated with several disorders, e.g., cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, von Gierke disease, and certain forms of diabetes mellitus. Single-gene defect diseases resulting in an inability to transport small molecules across membranes include, e.g., cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease (van't Hoff, W.G. (1996) Exp. Nephrol. 4:253-262; Talente, G.M. et al. (1994) Ann. Intern. Med. 120:218-226; and Chillon, M. et al. (1995) New Engl. J. Med. 332:1475-1480).

Human diseases caused by mutations in ion channel genes include disorders of skeletal muscle, cardiac muscle, and the central nervous system. Mutations in the pore-forming subunits of sodium and chloride channels cause myotonia, a muscle disorder in which relaxation after voluntary contraction is delayed. Sodium channel myotonias have been treated with channel blockers. Mutations in muscle

sodium and calcium channels cause forms of periodic paralysis, while mutations in the sarcoplasmic calcium release channel, T-tubule calcium channel, and muscle sodium channel cause malignant hyperthermia. Cardiac arrhythmia disorders such as the long QT syndromes and idiopathic ventricular fibrillation are caused by mutations in potassium and sodium channels (Cooper, E.C. and L.Y. Jan (1998) Proc. Natl. Acad. Sci. USA 96:4759-4766). All four known human idiopathic epilepsy genes code for ion channel proteins (Berkovic, S.F. and I.E. Scheffer (1999) Curr. Opin. Neurology 12:177-182). Other neurological disorders such as ataxias, hemiplegic migraine and hereditary deafness can also result from mutations in ion channel genes (Jen, J. (1999) Curr. Opin. Neurobiol. 9:274-280; Cooper, supra).

Ion channels have been the target for many drug therapies. Neurotransmitter-gated channels have been targeted in therapies for treatment of insomnia, anxiety, depression, and schizophrenia. Voltage-gated channels have been targeted in therapies for arrhythmia, ischemic stroke, head trauma, and neurodegenerative disease (Taylor, C.P. and L.S. Narasimhan (1997) Adv. Pharmacol. 39:47-98). Various classes of ion channels also play an important role in the perception of pain, and thus are potential targets for new analgesics. These include the vanilloid-gated ion channels, which are activated by the vanilloid capsaicin, as well as by noxious heat. Local anesthetics such as lidocaine and mexiletine which blockade voltage-gated Na⁺ channels have been useful in the treatment of neuropathic pain (Eglen, supra).

Ion channels in the immune system have recently been suggested as targets for immunomodulation. T-cell activation depends upon calcium signaling, and a diverse set of T-cell specific ion channels has been characterized that affect this signaling process. Channel blocking agents can inhibit secretion of lymphokines, cell proliferation, and killing of target cells. A peptide antagonist of the T-cell potassium channel Kv1.3 was found to suppress delayed-type hypersensitivity and allogenic responses in pigs, validating the idea of channel blockers as safe and efficacious immunosuppressants (Calahan, M.D. and K.G. Chandy (1997) Curr. Opin. Biotechnol. 8:749-756).

The discovery of new transporters and ion channels and the polynucleotides encoding them satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of transport, neurological, muscle, and immunological disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

SUMMARY OF THE INVENTION

The invention features purified polypeptides, transporters and ion channels, referred to collectively as "TRICH" and individually as "TRICH-1," "TRICH-2," "TRICH-3," "TRICH-4,"

“TRICH-5,” “TRICH-6,” “TRICH-7,” “TRICH-8,” “TRICH-9,” “TRICH-10,” “TRICH-11,”
“TRICH-12,” “TRICH-13,” “TRICH-14,” “TRICH-15,” “TRICH-16,” “TRICH-17,” “TRICH-18,”
“TRICH-19,” “TRICH-20,” “TRICH-21,” “TRICH-22,” “TRICH-23,” “TRICH-24,” “TRICH-25,”
“TRICH-26,” and “TRICH-27.” In one aspect, the invention provides an isolated polypeptide
5 comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence
selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence
having at least 90% sequence identity to an amino acid sequence selected from the group consisting of
SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group
consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected
10 from the group consisting of SEQ ID NO:1-27. In one alternative, the invention provides an isolated
polypeptide comprising the amino acid sequence of SEQ ID NO:1-27.

The invention further provides an isolated polynucleotide encoding a polypeptide comprising an
amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the
group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least
15 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-
27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of
SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group
consisting of SEQ ID NO:1-27. In one alternative, the polynucleotide encodes a polypeptide selected
from the group consisting of SEQ ID NO:1-27. In another alternative, the polynucleotide is selected
20 from the group consisting of SEQ ID NO:28-54.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter
sequence operably linked to a polynucleotide encoding a polypeptide comprising an amino acid
sequence selected from the group consisting of a) an amino acid sequence selected from the group
consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90%
25 sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c)
a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID
NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group
consisting of SEQ ID NO:1-27. In one alternative, the invention provides a cell transformed with the
recombinant polynucleotide. In another alternative, the invention provides a transgenic organism
30 comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide comprising an amino acid
sequence selected from the group consisting of a) an amino acid sequence selected from the group
consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90%
sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c)

a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27.

The invention further provides an isolated polynucleotide comprising a polynucleotide sequence selected from the group consisting of a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, b) a naturally occurring polynucleotide sequence having at least 90% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, c) a polynucleotide sequence complementary to a), d) a polynucleotide sequence complementary to b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide comprising a polynucleotide sequence selected from the group consisting of a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, b) a naturally occurring polynucleotide sequence having at least 90% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, c) a polynucleotide sequence complementary to a), d) a polynucleotide sequence complementary to b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said

target polynucleotide having a sequence of a polynucleotide comprising a polynucleotide sequence selected from the group consisting of a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, b) a naturally occurring polynucleotide sequence having at least 90% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, c) a polynucleotide sequence complementary to a), d) a polynucleotide sequence complementary to b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an agonist of a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide comprising an amino acid sequence selected from the group consisting

of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

The invention further provides a method of screening for a compound that specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide comprising an amino acid sequence selected from the group consisting of a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence selected from the group consisting of SEQ ID NO:28-54, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, and b) detecting altered expression of the target polynucleotide.

The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide comprising a polynucleotide sequence selected from the group consisting of i) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, ii) a naturally occurring polynucleotide sequence having at least 90% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, iii) a polynucleotide sequence complementary to i), iv) a polynucleotide sequence complementary to ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence selected from the group consisting of i) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, ii) a naturally occurring polynucleotide sequence having at least 90% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54, iii) a polynucleotide sequence complementary to i), iv) a polynucleotide sequence complementary to ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog for each polypeptide of the invention. The probability score for the match between each polypeptide and its GenBank homolog is also shown.

Table 3 shows structural features of each polypeptide sequence, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of each

polypeptide.

Table 4 lists the cDNA and genomic DNA fragments which were used to assemble each polynucleotide sequence, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for each polynucleotide of the invention.

5 Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

10

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will
15 be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so
20 forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described.
25 All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

30 "TRICH" refers to the amino acid sequences of substantially purified TRICH obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of TRICH. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other

compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

An "allelic variant" is an alternative form of the gene encoding TRICH. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding TRICH include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as TRICH or a polypeptide with at least one functional characteristic of TRICH. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding TRICH, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding TRICH. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent TRICH. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of TRICH is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence. Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of

TRICH. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

5 The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind TRICH polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or
10 synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

 The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to
15 immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

 The term "antisense" refers to any composition capable of base-pairing with the "sense"
20 (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense
25 molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

30 The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic TRICH, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

“Complementary” describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A “composition comprising a given polynucleotide sequence” and a “composition comprising a given amino acid sequence” refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding TRICH or fragments of TRICH may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

“Consensus sequence” refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (PE Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

“Conservative amino acid substitutions” are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
25	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
30	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
35	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr

Thr	Ser, Val
Trp	Phe, Tyr
Tyr	His, Phe, Trp
Val	Ile, Leu, Thr

5

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

10 A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one
15 biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

20 A "fragment" is a unique portion of TRICH or the polynucleotide encoding TRICH which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10,
25 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the
30 specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:28-54 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:28-54, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:28-54 is useful, for
35 example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:28-54 from related polynucleotide sequences. The precise length of a fragment of SEQ

ID NO:28-54 and the region of SEQ ID NO:28-54 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-27 is encoded by a fragment of SEQ ID NO:28-54. A fragment of SEQ ID NO:1-27 comprises a region of unique amino acid sequence that specifically identifies
5 SEQ ID NO:1-27. For example, a fragment of SEQ ID NO:1-27 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-27. The precise length of a fragment of SEQ ID NO:1-27 and the region of SEQ ID NO:1-27 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

10 A “full length” polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A “full length” polynucleotide sequence encodes a “full length” polypeptide sequence.

“Homology” refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

15 The terms “percent identity” and “% identity,” as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

20 Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191.

25 For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and “diagonals saved”=4. The “weighted” residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the “percent similarity” between aligned polynucleotide sequences.

30 Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at <http://www.ncbi.nlm.nih.gov/BLAST/>. The BLAST software suite includes various sequence analysis programs including “blastn,” that is used to align a known polynucleotide sequence with other

polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at <http://www.ncbi.nlm.nih.gov/gorf/bl2.html>. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 11

Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment

program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 3

Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for

annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 µg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 µg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C_0t or R_0t analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of

various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of TRICH which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of TRICH which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of TRICH. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of TRICH.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an TRICH may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of TRICH.

"Probe" refers to nucleic acid sequences encoding TRICH, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical

labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols. A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that

hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing
5 primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence.
10 This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, supra. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence.
15 Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be used to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated
20 regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent,
25 chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose
30 instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing TRICH, nucleic acids encoding TRICH, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For
5 example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free,
10 preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters,
15 chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

20 "Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection,
25 electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to
30 animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in

vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 95% or at least 98% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternative splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, or at least 98% or greater sequence identity over a certain defined length of one of the polypeptides.

THE INVENTION

The invention is based on the discovery of new human transporters and ion channels (TRICH), the polynucleotides encoding TRICH, and the use of these compositions for the diagnosis, treatment, or prevention of transport, neurological, muscle, and immunological disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the GenBank identification number (Genbank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability score for the match between each polypeptide and its GenBank homolog. Column 5 shows the annotation of the GenBank homolog along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of each of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Columns 1 and 2 list the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and the corresponding Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) for each polynucleotide of the invention. Column 3 shows the length of each polynucleotide sequence in basepairs. Column 4 lists fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:28-54 or that distinguish between SEQ ID NO:28-54 and related polynucleotide sequences. Column 5 shows identification numbers corresponding to cDNA sequences, coding sequences (exons) predicted from genomic DNA, and/or sequence assemblages

comprised of both cDNA and genomic DNA. These sequences were used to assemble the full length polynucleotide sequences of the invention. Columns 6 and 7 of Table 4 show the nucleotide start (5') and stop (3') positions of the cDNA and genomic sequences in column 5 relative to their respective full length sequences.

5 The identification numbers in Column 5 of Table 4 may refer specifically, for example, to Incyte cDNAs along with their corresponding cDNA libraries. For example, 6813453H1 is the identification number of an Incyte cDNA sequence, and ADRETUR01 is the cDNA library from which it is derived. Incyte cDNAs for which cDNA libraries are not indicated were derived from pooled cDNA libraries (e.g., 70207988V1). Alternatively, the identification numbers in column 5 may refer to
10 GenBank cDNAs or ESTs (e.g., g1947104) which contributed to the assembly of the full length polynucleotide sequences. Alternatively, the identification numbers in column 5 may refer to coding regions predicted by Genscan analysis of genomic DNA. For example, GNN.g6554406_006 is the identification number of a Genscan-predicted coding sequence, with g6554406 being the GenBank identification number of the sequence to which Genscan was applied. The Genscan-predicted coding
15 sequences may have been edited prior to assembly. (See Example IV.) Alternatively, the identification numbers in column 5 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. (See Example V.) Alternatively, the identification numbers in column 5 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon-stretching" algorithm. (See Example V.) In some cases, Incyte cDNA coverage redundant
20 with the sequence coverage shown in column 5 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotide sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to
25 assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses TRICH variants. A preferred TRICH variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the TRICH amino acid sequence, and which contains at least one functional or structural
30 characteristic of TRICH.

The invention also encompasses polynucleotides which encode TRICH. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:28-54, which encodes TRICH. The polynucleotide sequences of SEQ ID NO:28-54, as presented in the Sequence Listing, embrace the equivalent RNA sequences,

wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding TRICH. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding TRICH. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:28-54 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:28-54.

10 Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of TRICH.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding TRICH, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring TRICH, and all such variations are to be considered as being specifically disclosed.

15

Although nucleotide sequences which encode TRICH and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring TRICH under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding TRICH or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding TRICH and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

20

25

The invention also encompasses production of DNA sequences which encode TRICH and TRICH derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding TRICH or any fragment thereof.

30

Also encompassed by the invention are polynucleotide sequences that are capable of

hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:28-54 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) *Methods Enzymol.* 152:399-407; Kimmel, A.R. (1987) *Methods Enzymol.*

152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in

5 "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (PE Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (PE Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system 10 (PE Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

20 The nucleic acid sequences encoding TRICH may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) *PCR Methods Applic.* 2:318-322.)

25 Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) *Nucleic Acids Res.* 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. 30 (1991) *PCR Methods Applic.* 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) *Nucleic Acids Res.* 19:3055-3060).

Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo

Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a
5 GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library
10 does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-
15 specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, PE Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be
20 present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode TRICH may be cloned in recombinant DNA molecules that direct expression of TRICH, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent
25 amino acid sequence may be produced and used to express TRICH.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter TRICH-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic
30 oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent Number

5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of TRICH, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding TRICH may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.) Alternatively, TRICH itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp.55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (PE Biosystems). Additionally, the amino acid sequence of TRICH, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

In order to express a biologically active TRICH, the nucleotide sequences encoding TRICH or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences

encoding TRICH. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding TRICH. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding TRICH and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) *Results Probl. Cell Differ.* 20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding TRICH and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook, J. et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) Current Protocols in Molecular Biology, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding TRICH. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) *J. Biol. Chem.* 264:5503-5509; Engelhard, E.K. et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:3224-3227; Sandig, V. et al. (1996) *Hum. Gene Ther.* 7:1937-1945; Takamatsu, N. (1987) *EMBO J.* 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) *Proc. Natl. Acad. Sci. USA* 81:3655-3659; and Harrington, J.J. et al. (1997) *Nat. Genet.* 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) *Cancer Gen. Ther.* 5(6):350-356; Yu, M. et al. (1993) *Proc. Natl. Acad. Sci. USA* 90(13):6340-6344; Buller, R.M. et al. (1985) *Nature* 317(6040):813-815; McGregor, D.P. et al. (1994) *Mol. Immunol.* 31(3):219-226; and Verma, I.M. and N. Somia (1997) *Nature* 389:239-242.)

The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding TRICH. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding TRICH can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding TRICH into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of TRICH are needed, e.g. for the production of antibodies, vectors which direct high level expression of TRICH may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of TRICH. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast Saccharomyces cerevisiae or Pichia pastoris. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, supra; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of TRICH. Transcription of sequences encoding TRICH may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding TRICH may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses TRICH in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc.

Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of TRICH in cell lines is preferred. For example, sequences encoding TRICH can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in *tk⁻* and *apr⁻* cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *neo* confers resistance to the aminoglycosides neomycin and G-418; and *als* and *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β glucuronidase and its substrate β -glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding TRICH is inserted within a marker gene sequence, transformed cells containing

sequences encoding TRICH can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding TRICH under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

5 In general, host cells that contain the nucleic acid sequence encoding TRICH and that express TRICH may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

10 Immunological methods for detecting and measuring the expression of TRICH using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on TRICH is preferred, but a competitive binding
15 assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and
20 may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding TRICH include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding TRICH, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and
25 may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as
30 well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding TRICH may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing

polynucleotides which encode TRICH may be designed to contain signal sequences which direct secretion of TRICH through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding TRICH may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric TRICH protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of TRICH activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, *c-myc*, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, *c-myc*, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the TRICH encoding sequence and the heterologous protein sequence, so that TRICH may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled TRICH may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

TRICH of the present invention or fragments thereof may be used to screen for compounds

that specifically bind to TRICH. At least one and up to a plurality of test compounds may be screened for specific binding to TRICH. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of TRICH, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) Current Protocols in Immunology 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which TRICH binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express TRICH, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, Drosophila, or E. coli. Cells expressing TRICH or cell membrane fractions which contain TRICH are then contacted with a test compound and binding, stimulation, or inhibition of activity of either TRICH or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with TRICH, either in solution or affixed to a solid support, and detecting the binding of TRICH to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

TRICH of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of TRICH. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for TRICH activity, wherein TRICH is combined with at least one test compound, and the activity of TRICH in the presence of a test compound is compared with the activity of TRICH in the absence of the test compound. A change in the activity of TRICH in the presence of the test compound is indicative of a compound that modulates the activity of TRICH. Alternatively, a test compound is combined with an in vitro or cell-free system comprising TRICH under conditions suitable for TRICH activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of TRICH may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding TRICH or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES)

cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent Number 5,175,383 and U.S. Patent Number 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest
5 disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330).
10 Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding TRICH may also be manipulated in vitro in ES cells derived from
15 human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding TRICH can also be used to create "knockin" humanized animals
20 (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding TRICH is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a
25 mammal inbred to overexpress TRICH, e.g., by secreting TRICH in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

THERAPEUTICS

Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of TRICH and transporters and ion channels. Therefore, TRICH appears to play a
30 role in transport, neurological, muscle, and immunological disorders. In the treatment of disorders associated with increased TRICH expression or activity, it is desirable to decrease the expression or activity of TRICH. In the treatment of disorders associated with decreased TRICH expression or activity, it is desirable to increase the expression or activity of TRICH.

Therefore, in one embodiment, TRICH or a fragment or derivative thereof may be

administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrhythmia, tachyarrhythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathisia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses,

postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, and acid maltase deficiency (AMD, also known as Pompe's disease); and an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma.

In another embodiment, a vector capable of expressing TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified TRICH in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of TRICH may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those listed above.

In a further embodiment, an antagonist of TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH. Examples of such

disorders include, but are not limited to, those transport, neurological, muscle, and immunological disorders described above. In one aspect, an antibody which specifically binds TRICH may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express TRICH.

5 In an additional embodiment, a vector expressing the complement of the polynucleotide encoding TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate
10 therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

15 An antagonist of TRICH may be produced using methods which are generally known in the art. In particular, purified TRICH may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind TRICH. Antibodies to TRICH may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and
20 fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with TRICH or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to
25 increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to
30 TRICH have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of TRICH amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to TRICH may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) *Nature* 256:495-497; Kozbor, D. et al. (1985) *J. Immunol. Methods* 81:31-42; Cote, R.J. et al. (1983) *Proc. Natl. Acad. Sci. USA* 80:2026-2030; and Cole, S.P. et al. (1984) *Mol. Cell Biol.* 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) *Proc. Natl. Acad. Sci. USA* 81:6851-6855; Neuberger, M.S. et al. (1984) *Nature* 312:604-608; and Takeda, S. et al. (1985) *Nature* 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce TRICH-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) *Proc. Natl. Acad. Sci. USA* 88:10134-10137.)

Antibodies may also be produced by inducing in vivo production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) *Proc. Natl. Acad. Sci. USA* 86:3833-3837; Winter, G. et al. (1991) *Nature* 349:293-299.)

Antibody fragments which contain specific binding sites for TRICH may also be generated. For example, such fragments include, but are not limited to, $F(ab')_2$ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the $F(ab')_2$ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) *Science* 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between TRICH and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering TRICH epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for TRICH. Affinity is expressed as an association

constant, K_a , which is defined as the molar concentration of TRICH-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple TRICH epitopes, represents the average affinity, or avidity, of the antibodies for TRICH. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular TRICH epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10^9 to 10^{12} L/mole are preferred for use in immunoassays in which the TRICH-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10^6 to 10^7 L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of TRICH, preferably in active form, from the antibody (Catty, D. (1988) Antibodies. Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of TRICH-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding TRICH, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding TRICH. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding TRICH. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) *J. Allergy Clin. Immunol.* 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) *Blood* 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) *Pharmacol. Ther.* 63(3):323-347.) Other

gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

5 In another embodiment of the invention, polynucleotides encoding TRICH may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency
10 (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassemias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii)
15 express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA. 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides
20 brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in TRICH expression or regulation causes disease, the expression of TRICH from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

 In a further embodiment of the invention, diseases or disorders caused by deficiencies in
25 TRICH are treated by constructing mammalian expression vectors encoding TRICH and introducing these vectors by mechanical means into TRICH-deficient cells. Mechanical transfer technologies for use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-
30 217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

 Expression vectors that may be effective for the expression of TRICH include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF,

PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). TRICH may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β -actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. U.S.A. 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and Blau, H.M. supra), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding TRICH from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to TRICH expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding TRICH under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus *cis*-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. U.S.A. 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent Number 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4⁺ T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene

therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. U.S.A. 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In the alternative, an adenovirus-based gene therapy delivery system is used to deliver
5 polynucleotides encoding TRICH to cells which have one or more genetic abnormalities with respect to the expression of TRICH. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are
10 described in U.S. Patent Number 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

In another alternative, a herpes-based, gene therapy delivery system is used to deliver
15 polynucleotides encoding TRICH to target cells which have one or more genetic abnormalities with respect to the expression of TRICH. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing TRICH to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has
20 been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent Number 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent Number 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under
25 the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids
30 containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding TRICH to target cells. The biology of the prototypic alphavirus,

Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for TRICH into the alphavirus genome in place of the capsid-coding region results in the production of a large number of TRICH-coding RNAs and the synthesis of high levels of TRICH in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of TRICH into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding TRICH.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for

secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding TRICH. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding TRICH. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased TRICH expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding TRICH may be therapeutically useful, and in the treatment of disorders associated with decreased TRICH expression or activity, a compound which specifically promotes expression of the polynucleotide encoding TRICH may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary

library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding TRICH is exposed to at least one test compound thus obtained. The sample
5 may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding TRICH are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding TRICH. The amount of hybridization may be quantified, thus
10 forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression
15 system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide
20 sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient.
25 Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and
30 monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's

Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of TRICH, antibodies to TRICH, and mimetics, agonists, antagonists, or inhibitors of TRICH.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising TRICH or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, TRICH or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example TRICH or fragments thereof, antibodies of TRICH, and agonists, antagonists or inhibitors of TRICH, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose

lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD_{50}/ED_{50} ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED_{50} with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about $0.1 \mu\text{g}$ to $100,000 \mu\text{g}$, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

In another embodiment, antibodies which specifically bind TRICH may be used for the diagnosis of disorders characterized by expression of TRICH, or in assays to monitor patients being treated with TRICH or agonists, antagonists, or inhibitors of TRICH. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for TRICH include methods which utilize the antibody and a label to detect TRICH in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring TRICH, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of TRICH expression. Normal or standard values for TRICH expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to TRICH under conditions suitable for complex formation. The amount of standard complex formation may be

quantitated by various methods, such as photometric means. Quantities of TRICH expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding TRICH may be used for
5 diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of TRICH may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of TRICH, and to monitor regulation of TRICH levels during therapeutic intervention.

10 In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding TRICH or closely related molecules may be used to identify nucleic acid sequences which encode TRICH. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the
15 probe identifies only naturally occurring sequences encoding TRICH, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the TRICH encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:28-54 or from
20 genomic sequences including promoters, enhancers, and introns of the TRICH gene.

Means for producing specific hybridization probes for DNAs encoding TRICH include the cloning of polynucleotide sequences encoding TRICH or TRICH derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA
25 polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ^{32}P or ^{35}S , or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding TRICH may be used for the diagnosis of disorders associated with expression of TRICH. Examples of such disorders include, but are not limited to, a
30 transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral

- neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrhythmia, tachyarrhythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis,
- 5 infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia,
- 10 hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's
- 15 disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and
- 20 Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other
- 25 neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathisia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration,
- 30 and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's
- 35 syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and

myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, and acid maltase deficiency (AMD, also known as Pompe's disease); and an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma. The polynucleotide sequences encoding TRICH may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered TRICH expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding TRICH may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding TRICH may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding TRICH in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of TRICH, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding TRICH, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with

values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

5 Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

10 With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further
15 progression of the cancer.

 Additional diagnostic uses for oligonucleotides designed from the sequences encoding TRICH may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding TRICH, or a fragment of a polynucleotide complementary to the polynucleotide encoding TRICH, and
20 will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

 In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH may be used to detect single nucleotide polymorphisms (SNPs). SNPs are
25 substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal
30 tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSSCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed *in silico* SNP

(isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the
5 alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of TRICH include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et
10 al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the
15 polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor
20 progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

25 In another embodiment, TRICH, fragments of TRICH, or antibodies specific for TRICH may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of
30 gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent Number 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of

transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

5 Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of
10 pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a
15 signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the
20 rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released
25 February 29, 2000, available at <http://www.niehs.nih.gov/oc/news/toxchip.htm>.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated
30 biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for TRICH to quantify the levels of TRICH expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lueking, A. et al. (1999) *Anal. Biochem.* 270:103-111; Mendoze, L.G. et al. (1999) *Biotechniques* 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) *Electrophoresis* 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference
5 in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated
10 with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g.,
15 Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in DNA Microarrays: A Practical Approach, M. Schena, ed. (1999)
20 Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding TRICH may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members
25 of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat.
30 Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding TRICH on a physical
5 map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may
10 reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g.,
15 Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, TRICH, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug
20 screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between TRICH and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT
25 application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with TRICH, or fragments thereof, and washed. Bound TRICH is then detected by methods well known in the art. Purified TRICH can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a
30 solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding TRICH specifically compete with a test compound for binding TRICH. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with TRICH.

In additional embodiments, the nucleotide sequences which encode TRICH may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

5 Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications and publications, mentioned above and below, in
10 particular U.S. Ser. No. 60/172,000, U.S. Ser. No. 60/176,083, U.S. Ser. No. 60/177,332, U.S. Ser. No. 60/178,572, U.S. Ser. No. 60/179,758, and U.S. Ser. No. 60/181,625, are expressly incorporated by reference herein.

EXAMPLES

15 I. Construction of cDNA Libraries

Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA) and shown in Table 4, column 5. Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a
20 suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated
25 using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA
30 libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERScript plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the

appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g.,

5 PBLUESCRIPT plasmid (Stratagene), PSORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), or pINCY (Incyte Genomics, Palo Alto CA). Recombinant plasmids were transformed into competent *E. coli* cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5 α , DH10B, or ElectroMAX DH10B from Life Technologies.

II. Isolation of cDNA Clones

10 Plasmids obtained as described in Example I were recovered from host cells by *in vivo* excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid

15 purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-

20 well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows.

25 Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (PE Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI

30 PRISM BIGDYE Terminator cycle sequencing ready reaction kit (PE Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (PE Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA

sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families.

See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMR. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length.

Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences which were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and

polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:28-54. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 4.

IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative transporters and ion channels were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode transporters and ion channels, the encoded polypeptides were analyzed by querying against PFAM models for transporters and ion channels. Potential transporters and ion channels were also identified by homology to Incyte cDNA sequences that had been annotated as transporters and ion channels. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data

"Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in

the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpr public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

"Stretched" Sequences

Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of TRICH Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:28-54 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:28-54 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment

of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, or human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Génethon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (<http://www.ncbi.nlm.nih.gov/genemap/>), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

VII. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, supra, ch. 7; Ausubel (1995) supra, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

$$\frac{\text{BLAST Score} \times \text{Percent Identity}}{5 \times \text{minimum} \{ \text{length}(\text{Seq. 1}), \text{length}(\text{Seq. 2}) \}}$$

25

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter

of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding TRICH are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding TRICH. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

VIII. Extension of TRICH Encoding Polynucleotides

Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg^{2+} , $(NH_4)_2SO_4$, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer

pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; 5 Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 µl PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 µl of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II 10 (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 µl to 10 µl aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and 15 sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, 20 and transfected into competent *E. coli* cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 25 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethylsulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC 30 DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (PE Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:28-54 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments.

- 5 Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [γ - 32 P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10^7 counts per
10 minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

- The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16
15 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

X. Microarrays

- 20 The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, supra), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena (1999), supra). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure
25 analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol.
30 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the

biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection.

After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorption and mass spectrometry may be used for detection of hybridization. The degree of

5 complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)⁺ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)⁺ RNA sample is
10 reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/ μ l oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/ μ l RNase inhibitor, 500 μ M dATP, 500 μ M dGTP, 500 μ M dTTP, 40 μ M dCTP, 40 μ M dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)⁺ RNA with
15 GEMBRIGHT kits (Incyte). Specific control poly(A)⁺ RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37° C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85° C to stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc.
20 (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μ l 5X SSC/0.2% SDS.

Microarray Preparation

25 Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia
30 Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and

coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522, incorporated herein by reference. 1 µl of the array element DNA, at an average concentration of 100 ng/µl, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

Hybridization

Hybridization reactions contain 9 µl of sample mixture consisting of 0.2 µg each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 µl of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

XI. Complementary Polynucleotides

Sequences complementary to the TRICH-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring TRICH. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of TRICH. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the TRICH-encoding transcript.

XII. Expression of TRICH

Expression and purification of TRICH is achieved using bacterial or virus-based expression systems. For expression of TRICH in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the *trp-lac (tac)* hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory

element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express TRICH upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of TRICH in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding TRICH by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, TRICH is synthesized as a fusion protein with, e.g., glutathione S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from TRICH at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified TRICH obtained by these methods can be used directly in the assays shown in Examples XVI, XVII, and XVIII, where applicable.

XIII. Functional Assays

TRICH function is assessed by expressing the sequences encoding TRICH at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the

recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of TRICH on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding TRICH and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding TRICH and other genes of interest can be analyzed by northern analysis or microarray techniques.

XIV. Production of TRICH Specific Antibodies

TRICH substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) *Methods Enzymol.* 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the TRICH amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (PE Biosystems) using Fmoc chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-TRICH

activity by, for example, binding the peptide or TRICH to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XV. Purification of Naturally Occurring TRICH Using Specific Antibodies

Naturally occurring or recombinant TRICH is substantially purified by immunoaffinity chromatography using antibodies specific for TRICH. An immunoaffinity column is constructed by covalently coupling anti-TRICH antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing TRICH are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of TRICH (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/TRICH binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and TRICH is collected.

XVI. Identification of Molecules Which Interact with TRICH

Molecules which interact with TRICH may include transporter substrates, agonists or antagonists, modulatory proteins such as G $\beta\gamma$ proteins (Reimann, *supra*) or proteins involved in TRICH localization or clustering such as MAGUKs (Craven, *supra*). TRICH, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH complex are assayed. Data obtained using different concentrations of TRICH are used to calculate values for the number, affinity, and association of TRICH with the candidate molecules.

Alternatively, proteins that interact with TRICH are isolated using the yeast 2-hybrid system (Fields, S. and O. Song (1989) Nature 340:245-246). TRICH, or fragments thereof, are expressed as fusion proteins with the DNA binding domain of Gal4 or lexA and potential interacting proteins are expressed as fusion proteins with an activation domain. Interactions between the TRICH fusion protein and the reconstitutes a transactivation function that is observed by expression of a reporter gene. Yeast 2-hybrid systems are commercially available, and methods for use of the yeast 2-hybrid system with ion channel proteins are discussed in Niethammer, M. and M. Sheng (1998, Meth. Enzymol. 293:104-122).

TRICH may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

Potential TRICH agonists or antagonists may be tested for activation or inhibition of TRICH

ion channel activity using the assays described in section XVIII.

XVII. Demonstration of TRICH Activity

Ion channel activity of TRICH is demonstrated using an electrophysiological assay for ion conductance. TRICH can be expressed by transforming a mammalian cell line such as COS7, HeLa
5 or CHO with a eukaryotic expression vector encoding TRICH. Eukaryotic expression vectors are commercially available, and the techniques to introduce them into cells are well known to those skilled in the art. A second plasmid which expresses any one of a number of marker genes, such as β -galactosidase, is co-transformed into the cells to allow rapid identification of those cells which have taken up and expressed the foreign DNA. The cells are incubated for 48-72 hours after transformation
10 under conditions appropriate for the cell line to allow expression and accumulation of TRICH and β -galactosidase.

Transformed cells expressing β -galactosidase are stained blue when a suitable colorimetric substrate is added to the culture media under conditions that are well known in the art. Stained cells are tested for differences in membrane conductance by electrophysiological techniques that are well
15 known in the art. Untransformed cells, and/or cells transformed with either vector sequences alone or β -galactosidase sequences alone, are used as controls and tested in parallel. Cells expressing TRICH will have higher anion or cation conductance relative to control cells. The contribution of TRICH to conductance can be confirmed by incubating the cells using antibodies specific for TRICH. The antibodies will bind to the extracellular side of TRICH, thereby blocking the pore in the ion channel,
20 and the associated conductance.

Alternatively, ion channel activity of TRICH is measured as current flow across a TRICH-containing Xenopus laevis oocyte membrane using the two-electrode voltage-clamp technique (Ishi et al., supra; Jegla, T. and L. Salkoff (1997) J. Neurosci. 17:32-44). TRICH is subcloned into an appropriate Xenopus oocyte expression vector, such as pBF, and 0.5-5 ng of mRNA is injected into
25 mature stage IV oocytes. Injected oocytes are incubated at 18°C for 1-5 days. Inside-out macropatches are excised into an intracellular solution containing 116 mM K-gluconate, 4 mM KCl, and 10 mM Hepes (pH 7.2). The intracellular solution is supplemented with varying concentrations of the TRICH mediator, such as cAMP, cGMP, or Ca^{+2} (in the form of CaCl_2), where appropriate. Electrode resistance is set at 2-5 M Ω and electrodes are filled with the intracellular solution lacking
30 mediator. Experiments are performed at room temperature from a holding potential of 0 mV. Voltage ramps (2.5 s) from -100 to 100 mV are acquired at a sampling frequency of 500 Hz. Current measured is proportional to the activity of TRICH in the assay.

Transport activity of TRICH is assayed by measuring uptake of labeled substrates into Xenopus laevis oocytes. Oocytes at stages V and VI are injected with TRICH mRNA (10 ng per

oocyte) and incubated for 3 days at 18°C in OR2 medium (82.5mM NaCl, 2.5 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 1mM Na₂HPO₄, 5 mM Hepes, 3.8 mM NaOH, 50µg/ml gentamycin, pH 7.8) to allow expression of TRICH. Oocytes are then transferred to standard uptake medium (100mM NaCl, 2 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 10 mM Hepes/Tris pH 7.5). Uptake of various substrates (e.g., amino acids, sugars, drugs, ions, and neurotransmitters) is initiated by adding labeled substrate (e.g. radiolabeled with ³H, fluorescently labeled with rhodamine, etc.) to the oocytes. After incubating for 30 minutes, uptake is terminated by washing the oocytes three times in Na⁺-free medium, measuring the incorporated label, and comparing with controls. TRICH activity is proportional to the level of internalized labeled substrate.

ATPase activity associated with TRICH can be measured by hydrolysis of radiolabeled ATP-³²P, separation of the hydrolysis products by chromatographic methods, and quantitation of the recovered ³²P using a scintillation counter. The reaction mixture contains ATP-³²P and varying amounts of TRICH in a suitable buffer incubated at 37°C for a suitable period of time. The reaction is terminated by acid precipitation with trichloroacetic acid and then neutralized with base, and an aliquot of the reaction mixture is subjected to membrane or filter paper-based chromatography to separate the reaction products. The amount of ³²P liberated is counted in a scintillation counter. The amount of radioactivity recovered is proportional to the ATPase activity of TRICH in the assay.

XVIII. Identification of TRICH Agonists and Antagonists

TRICH is expressed in a eukaryotic cell line such as CHO (Chinese Hamster Ovary) or HEK (Human Embryonic Kidney) 293. Ion channel activity of the transformed cells is measured in the presence and absence of candidate agonists or antagonists. Ion channel activity is assayed using patch clamp methods well known in the art or as described in Example XVII. Alternatively, ion channel activity is assayed using fluorescent techniques that measure ion flux across the cell membrane (Velicelebi, G. et al. (1999) Meth. Enzymol. 294:20-47; West, M.R. and C.R. Molloy (1996) Anal. Biochem. 241:51-58). These assays may be adapted for high-throughput screening using microplates. Changes in internal ion concentration are measured using fluorescent dyes such as the Ca²⁺ indicator Fluo-4 AM, sodium-sensitive dyes such as SBFI and sodium green, or the Cl⁻ indicator MQAE (all available from Molecular Probes) in combination with the FLIPR fluorimetric plate reading system (Molecular Devices). In a more generic version of this assay, changes in membrane potential caused by ionic flux across the plasma membrane are measured using oxonyl dyes such as DiBAC₄ (Molecular Probes). DiBAC₄ equilibrates between the extracellular solution and cellular sites according to the cellular membrane potential. The dye's fluorescence intensity is 20-fold greater when bound to hydrophobic intracellular sites, allowing detection of DiBAC₄ entry into the cell (Gonzalez, J.E. and P.A. Negulescu (1998) Curr. Opin. Biotechnol. 9:624-631). Candidate agonists or antagonists may be

selected from known ion channel agonists or antagonists, peptide libraries, or combinatorial chemical libraries.

Various modifications and variations of the described methods and systems of the invention will
5 be apparent to those skilled in the art without departing from the scope and spirit of the invention.
Although the invention has been described in connection with certain embodiments, it should be
understood that the invention as claimed should not be unduly limited to such specific embodiments.
Indeed, various modifications of the described modes for carrying out the invention which are obvious
to those skilled in molecular biology or related fields are intended to be within the scope of the following
10 claims.

Table 1

Incyte Project ID	Polypeptide SEQ ID NO:	Incyte Polypeptide ID	Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID
1416107	1	1416107CD1	28	1416107CB1
1682513	2	1682513CD1	29	1682513CB1
2446438	3	2446438CD1	30	2446438CB1
2817822	4	2817822CD1	31	2817822CB1
4009329	5	4009329CD1	32	4009329CB1
6618083	6	6618083CD1	33	6618083CB1
7472002	7	7472002CD1	34	7472002CB1
1812692	8	1812692CD1	35	1812692CB1
3232992	9	3232992CD1	36	3232992CB1
3358383	10	3358383CD1	37	3358383CB1
4250091	11	4250091CD1	38	4250091CB1
70064803	12	70064803CD1	39	70064803CB1
70356768	13	70356768CD1	40	70356768CB1
5674114	14	5674114CD1	41	5674114CB1
1254635	15	1254635CD1	42	1254635CB1
1670595	16	1670595CD1	43	1670595CB1
1859560	17	1859560CD1	44	1859560CB1
5530164	18	5530164CD1	45	5530164CB1
139115	19	139115CD1	46	139115CB1
1702940	20	1702940CD1	47	1702940CB1
1703342	21	1703342CD1	48	1703342CB1
1727529	22	1727529CD1	49	1727529CB1
2289333	23	2289333CD1	50	2289333CB1
2720354	24	2720354CD1	51	2720354CB1
3038193	25	3038193CD1	52	3038193CB1
3460979	26	3460979CD1	53	3460979CB1
7472200	27	7472200CD1	54	7472200CB1

Table 2

Polypeptide SEQ ID NO:	Incyte Polypeptide ID	GenBank ID NO:	Probability Score	GenBank Homolog
1	1416107CD1	g7018605	1.9e-302	Glucose transporter [Rattus norvegicus] (Ibberson, M. et al. (2000) J. Biol. Chem. 275:4607-4612)
2	1682513CD1	g5263196	1.4e-153	Stretch-inhibitable nonselective channel (SIC) [Rattus norvegicus] (Cloning of a stretch-inhibitable nonselective cation channel. J Biol Chem. 1999 Mar 5;274(10):6330-6335)
3	2446438CD1	g4589141	0	Vanilloid receptor-like protein 1 [Homo sapiens] (A capsaicin-receptor homologue with a high threshold for noxious heat. Nature 1999 398:436-441)
5	4009329CD1	g3873983	1.9e-64	Similar to Na+/Ca+, K+ antiporter [C. elegans]
6	6618083CD1	g9230651	4.7e-268	Facilitative glucose transporter family member GLUT9 [Homo sapiens] (Phay, J.E. et al. (2000) Genomics 66:217-220)
7	7472002CD1	g433960	0	Aorta CNG channel (rACNG) [Oryctolagus cuniculus] (Primary structure and functional expression of a cyclic nucleotide-gated channel from rabbit aorta. FEBS Lett. 1993 Aug 23;329(1-2):134-138)
8	1812692CD1	g3928756	4.5e-48	Transient receptor potential channel 7 [Homo sapiens] (Nagamine, K. et al. (1998) Molecular cloning of a novel putative Ca2+ channel protein (TRPC7) highly expressed in brain)
9	3232992CD1	g3874275	3.2e-70	Similarity to yeast low-affinity glucose transporter HXT4 [Caenorhabditis elegans]
10	3358383CD1	g3004482	1.4e-163	Putative integral membrane transport protein [Rattus norvegicus] (Schomig, E. et al. (1998) Molecular cloning and characterization of two novel transport proteins from rat kidney. FEBS Lett. 425:79-86)
11	4250091CD1	g3880445	5.7e-16	VM106R.1 (similar to K+ channel tetramerisation domain) [Caenorhabditis elegans]
12	70064803CD1	g3874275	7.0e-84	Similarity to yeast low-affinity glucose transporter HXT4 [Caenorhabditis elegans]
13	70356768CD1	g183298	4.1e-54	GLUT5 protein [Homo sapiens] (Kayano, Y. et al. (1990) Human facilitative glucose transporters. Isolation, functional characterization, and gene localization of cDNAs encoding an isoform (GLUT5) expressed in small intestine, kidney, muscle, and adipose tissue and an unusual glucose transporter pseudogene-like sequence (GLUT6). J. Biol. Chem. 265:13276-13282)

Table 2 (cont.)

Polypeptide SEQ ID NO:	Incyte Polypeptide ID	GenBank ID NO:	Probability Score	GenBank Homolog
14	5674114CD1	g5771352	1.3e-238	Inward rectifier potassium channel Kir2.4 [Homo sapiens] (Topert, C. et al. (1998) Kir2.4: a novel K ⁺ inward rectifier channel associated with motoneurons of cranial nerve nuclei. J. Neurosci. 18:4096-4105)
15	1254635CD1	g3953533	1.7e-210	Inwardly rectifying potassium channel Kir5.1 [Mus musculus] (Mouri, T. et al. (1998) Assignment of mouse inwardly rectifying potassium channel Kcnj16 to the distal region of mouse chromosome 11. Genomics 54:181-182)
16	1670595CD1	g9502260	2.3e-146	Cation-chloride cotransporter-interacting protein [Homo sapiens] (Caron, L. et al. (2000) J. Biol. Chem. 275:32027-32036)
17	1859560CD1	g5834394	1.4e-101	Sulfate transporter [Drosophila melanogaster]
18	5530164CD1	g4903004	3.5e-20	UDP-N-acetylglucosamine transporter [Homo sapiens] (Ishida, N. et al. (1999) Molecular cloning and functional expression of the human golgi UDP-N-acetylglucosamine transporter. J. Biochem. 126:68-77.)
19	139115CD1	g8131858	1.5e-49	Putative thymic stromal co-transporter TSCOT [Mus musculus] (Kim, M.G. et al. (2000) J. Immunol. 164:3185-3192)
20	1702940CD1	g5725224	2.5e-143	bK212A2.2 (similar to apolipoprotein L) [Homo sapiens]
21	1703342CD1	g6003536	8.1e-06	Calcium channel alpha-1 subunit [Bdellovira candida]
22	1727529CD1	g4529890	0.0	NG22 [Homo sapiens]
23	2289333CD1	g4539333	5.6e-35	Putative amino acid transport protein [Arabidopsis thaliana]
24	2720354CD1	g3875242	4.3e-38	Similar to mitochondrial carrier protein [Caenorhabditis elegans]
26	3460979CD1	g1931644	4.7e-08	Membrane protein PTM1 precursor isolog (putative major facilitator superfamily transporter) [Arabidopsis thaliana]
27	7472200CD1	g2811254	2.8e-21	Amiloride-sensitive Na ⁺ channel [Drosophila melanogaster] (Adams, C.M. et al. (1998) J. Cell Biol. 140:143-152)

Table 3

SEQ ID NO:	Incyte Polypeptide ID	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
1	1416107CD1	477	S99 T2 T281 S430 T205	N349	Sugar transporter domain: A29-F474 Sugar transport protein signatures: V108-L174, L293-S350, G41-I51, L124-V143, Q267-F277, V375-L396, S398-T410 Glucose transporter signatures: F257-Y278, V375-S398, G439-V459 Transmembrane domains: I259-A279, L293-M313, L320-Y339, Y438-F457	MOTIFS HMMER-PFAM BLIMPS-BLOCKS ProfileScan BLIMPS-PRINTS HMMER
2	1682513CD1	498	S47 T131 S286 T367 S463 T67 S315 S382	N278 N411 N429	Glucose transporter signature: P319-N339 Transmembrane domains: A95-Y117, V142-F160, L178-Y201, A200-F219, F244-L263, P319-N339	MOTIFS BLIMPS-PRINTS HMMER
3	2446438CD1	764	T64 T329 S23 S67 T106 S268 S339 S348 S353 S464 S468 S667 T692 S697 T720 T101 T115 S325 T414 Y110 Y227 Y333	N570	Ankyrin repeat: R162-C194; F208-S243 Q293-F328 Transmembrane domains: L386-F405, I463-V486, F538-S557, L623-I642	MOTIFS HMMER-PFAM HMMER
4	2817822CD1	255	T30 S167 T33 T71 S23 T50 S134 T162 T238		Potassium channel signature: R76-T95	MOTIFS BLIMPS-PRINTS
5	4009329CD1	584	S258 S321 T70 S271 S273 S468 S514 S62 T132	N60 N125	Signal peptide: M1-G29 Sodium/calcium exchanger protein domain: I113-Q252, L431-F576 Transmembrane domains: T101-F121, T166-I189, L234-Y251, Y382-A402, F492-R513, L560-M584	MOTIFS HMMER HMMER-PFAM HMMER

Table 3 (cont.)

SEQ ID NO:	Incyte Polypeptide ID	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
6	6618083CD1	416	T111 S4 S164 S274 T374 S16 S226 S269	N45 N61 N410	Signal peptide: M1-G37 Sugar transporter domain: A30-E416 Sugar transport proteins signatures: A123-L189, S39-V49, I139-M158, Y298-F308 Glucose transporter signatures: V288-Y309, I356-Q376 Transmembrane domains: V83-V99, I356-L375	MOTIFS SPScan HMMER HMMER-PFAM BLIMPS-BLOCKS ProfileScan BLIMPS-PRINTS HMMER
7	7472002CD1	664	S402 S40 S46 S93 T107 T313 T337 T381 T422 S476 S552 T591 T606 T634 T2 S35 T124 T208 T418 T448 Y648	N311 N379	Transmembrane region, cyclic nucleotide gated channel: Y215-I440 Cyclic nucleotide binding domain: K469-D565, A460-S476, G478-V501, G516-L525 Transmembrane domain: Y350-I375	MOTIFS HMMER-PFAM BLIMPS-BLOCKS HMMER
8	1812692CD1	242	T95 S35 S37 T124 S204 S221 S2 S9 S20 T21 S86 Y36	N15 N84	Protein melastatin chromosome transmembrane PD018035: Y117-I227	BLAST- PRODOM HMMER
9	3232992CD1	398	T307 S338 S100 T133 T241 T303 S377 S395	N161	Transmembrane domains: A217-Q242, L247-F264, L350-F368 Sugar transport proteins signature: L45-G94, V30-I96	MOTIFS BLIMPS-BLOCKS ProfileScan HMMER
10	3358383CD1	553	S337 S352 S409 T58 S60 S109 T133 S337 T433 T527 S167 S201 T226 S282 T323 T405	N39 N56 N62 N102 N107 N473	Transmembrane domains: F204-A222, M470-Y493, I500-T519 glpT family of transporters: V151-D168 Organic transport protein, renal anion transporter, cationic kidney specific solute PD151320: N102-L144	MOTIFS BLIMPS-BLOCKS BLAST- PRODOM

Table 3 (cont.)

SEQ ID NO:	Incyte Polypeptide ID	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
11	4250091CD1	213	S2 S93 S172 S184 T17 T22 T137 S210 Y89	N188	Potassium channel signature: Q48-T67	MOTIFS BLIMPS- PRINTS
12	70064803CD1	476	T365 S11 S364 S453 S292 T361 S390 S466		Transmembrane domains: V222-R239, G327-V350, M413-F432 Sugar transport proteins signature: L153-G202, V138-I204	HMMER MOTIFS BLIMPS- BLOCKS ProfileScan
13	70356768CD1	246	S100 S238 S118 S215	N34 N50	Signal peptide: M1-G27 Sugar transport proteins signature: I127-G176, A112-V178, T28-I38, M128-M147, M133-R158	MOTIFS SPScan HMMER BLIMPS- BLOCKS ProfileScan BLIMPS- PRINTS
14	5674114CD1	436	S11 T80 S154 S340 S362 T263 S376 S422 Y47	N195	Transmembrane domain: M163-L181 Sugar transport proteins: DM00135 P46408 : A112-C229 Inward rectifier potassium channel domain: V53-L394, R72-L118, P126-Q169, C170-V199, A204-Q238, D296-Y346, T358-E368 Transmembrane domain: W88-L114 Inward rectifier potassium channel: DM00448 P52188 : N34-A395 Inward rectifier potassium channel: PD001103: V53-Q372 KIR2.4 protein: PD124342: A373-P436 PD063376: M1-F52	HMMER BLAST-DOMO MOTIFS HMMER-PFAM BLIMPS-PFAM HMMER BLAST-DOMO BLAST- PRODOM

Table 3 (cont.)

SEQ ID NO:	Incyte Polypeptide ID	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
15	1254635CD1	453	S408 T16 T99 S416 S29 T209 T216 T250 S375		Signal peptide: M1-A32 Transmembrane domains: F109-A131, V182-I200 Inward rectifier potassium channel domain: L72-T399 Inward rectifier potassium ion channel subfamily PD001103: K74-K403 Voltage-gated inward rectifier potassium channel BIR9, KIR5.1, transmembrane PD063375: M36-H73 Inward rectifier potassium channel: DM00448 P52185 27-380: K64-L379 Inward rectifier potassium channel signature: S91-L137, P145-Q188, S189-R218, A223-R257, N310-C360, A371-W381 Transmembrane domains: L37-L58, A74-I93, L134-Y154, F159-V179 Sensitive cotransporter, chloride, sodium: DM01178 S06903 1-128: A32-L153	SPScan HMMER HMMER-PFAM BLAST-PRODOM BLAST-PRODOM BLAST-DOMO BLIMPS-PFAM HMMER BLAST-DOMO
16	1670595CD1	299	S38 T211 S263 S33 T187	N193 N208		

Table 3 (cont.)

SEQ ID NO:	Incyte Polypeptide ID	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
17	1859560CD1	606	T96 T116 S298 S571 T572 S595 T245	N294	Transmembrane domains: L45-I63, T396-T421 Sulfate transporter family domain: M137-A468 Sulfate transporters protein signature: A54-I107, L125-L176 Sulfate transport protein, transmembrane, permease PD001255: M137-L465 Sulfate transport protein, transmembrane, permease PD001121: L30-G143 Sulfate transporter: DM01229 S64926 69-531: L30-W428 Sulfate transporters motif: P77-R98	HMMER HMMER-PFAM BLIMPS- BLOCKS BLAST- PRODOR BLAST-DOMO MOTIFS
18	5530164CD1	324	S2 S139 Y115	N99 N100 N101 N232	Transmembrane domains: A145-V163, L302-L319	HMMER
19	139115CD1	445	S54 S32 S77 S217 S424 S438 T150 S237 S443 Y23	N22 N30 N37 N127 N213 N235	Transmembrane domains: W182-F200, F242-I261, Y283-F302 Sugar transporter motif: L75-S91 Glucose transporter signature: W182-L202	HMMER MOTIFS BLIMPS- PRINTS
20	1702940CD1	337	T30 S167 T208 S306		Apolipoprotein L precursor, lipid transport, glycoprotein, signal, DJ6802.1 PD042084: M1-D336	BLAST- PRODOR
21	1703342CD1	273	T3 S63 T222 S248 S250 T10 S98 S219 S224		Transmembrane domains: F101-A118, M142-F161, F170-I186, I202-I218 Ion transport protein domain: L95-L269 (Score: -132.1, E-value: 0.72)	HMMER HMMER-PFAM

Table 3 (cont.)

SEQ ID NO:	Incyte Polypeptide ID	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
22	1727529CD1	710	S31 S102 S119 T135 S304 S22 S218 S430 S431 T494 S573 S619 Y13	N29 N69 N155 N197 N298 N393 N405 N416 N678	Transmembrane domains: C38-V58, V241-L266, W309-V326, F356-T375, F440-L458, T499-I522, L598-F618, I645-V663 ABC 3 transport family: S228-Q427 (Score: -182.9, E-value: 2.1) Anion exchanger signature: A311-L330	HMMER
23	2289333CD1	476	T97 T7 S8 S125 T443 S272 S322 T351 T451 Y184	N166 N169 N212 N425 N467	Transmembrane domains: L54-I81, V127-F145, Y184-S208, L279-G297, I331-K357, I426-T451 Transmembrane amino acid transporter protein domain: A55-F436 Amino acid transporter protein, permease, transmembrane, putative proline PD001875: D27-I337 Signal peptide: M1-G15 Mitochondrial carrier proteins domain: S25-L109, L122-T202 Mitochondrial energy transfer proteins signature: L128-Q152 Mitochondrial energy transfer proteins signature: L27-I75, V123-Q171	BLIMPS- PRODOR HMMER HMMER-PFAM BLAST- PRODOR SPScan HMMER-PFAM
24	2720354CD1	237	T17 T64 S172		Mitochondrial carrier proteins signature: G87-D107, V136-D154 Adenine nucleotide translocator 1 signature: R63-V84, E176-R191 Mitochondrial carrier proteins motifs: P46-L54, P143-L151 Transport protein, transmembrane, inner mitochondrial, ADP/ATP: PD000117: L31-F200 Mitochondrial energy transfer protein: DM00026 P38087 243-325: L128-Y209	BLIMPS- PRINTS BLIMPS- PRINTS MOTIFS BLAST- PRODOR BLAST-DOMO

Table 3 (cont.)

SEQ ID NO:	Incyte Polypeptide ID	Amino Acid Residues	Potential Phosphorylation Sites	Potential Glycosylation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
25	3038193CD1	345	T204 T251 S57 S243 T263 T308 S340	N246	Transmembrane domains: L67-L95, I134-I156, I224-F242 Sodium bile acid symporter family: Y44-P212 (Score: -7.0, E-value: 9.0e-4) Phosphate transporter signature: F153-G171	HMMER HMMER-PFAM
26	3460979CD1	521	S115 T184 S75 T93 S100 S126 S128 S134 S148 S183 S213 S256 S363 S389 S430 S510 T171 S180 S235 T247 S422 Y506	N70 N169 N211	Transmembrane domains: L265-L284, I335-I361 Protein precursor PTM1, transmembrane, signal PD014374: G219-E517 (P-value: 7.1e-07)	BLIMPS- PRODROM HMMER BLAST- PRODROM
27	7472200CD1	555	T43 S56 T92 T148 T298 S423 S468 S20 S52 S82 S96 T184 S208 S252 S393	N132 N175 N311 N361 N421	Amiloride-sensitive sodium channel alpha subunit signature PR01078: Y102-N118, Y342-Q353, Q353-P370, Q388-N404, G455-E471 Transmembrane domain: V452-F475 Amiloride-sensitive sodium channel ASC: F38-L476 Amiloride-sensitive sodium channel proteins BL01206: R37-L47, Y342-F368, L427-L472	BLIMPS- PRINTS HMMER HMMER-PFAM BLIMPS- BLOCKS

Table 4

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragments	Sequence Fragments	5' Position	3' Position
28	1416107CB1	2080	1-109, 1901-2080, 1363-1446	g1941704	116	609
				6813453H1 (ADRETUR01)	319	870
				6605280H1 (UTREDIT07)	820	1447
				881845R1 (THYRNOT02)	889	1479
				1416107F6 (BRAINOT12)	1348	1896
				1416107T6 (BRAINOT12)	1579	2080
				71826604V1	1563	1974
				7448905T1 (BRAYDIN03)	1578	2050
				6300413H1 (UTREDIT07)	423	752
				71805807V1	750	1580
				71827149V1	1584	2080
				71807187V1	701	1241
				6813453R6 (ADRETUR01)	1	312
29	1682513CB1	2128	1-1535, 1560-1581	70207988V1	1	469
				70213506V1	394	872
				70211216V1	489	985
				70210573V2	852	1468
				70207907V1	988	1512
				70210540V2	1357	1948
				2866122T6 (KIDNNOT20)	1548	2108
				70211461V1	1597	2128
				5073532H2 (COLCTUT03)	1	311
				6309494H1 (NERDIDN03)	260	812
30	2446438CB1	2825	1-65, 2000-2202, 999-1820	6268005H1 (MCLDTXN03)	344	996
				70382927D1	996	1525
				70386205D1	1469	2075
				1798255F6 (COLNNOT27)	1632	2194
				1562088F6 (SPLNNOT04)	2178	2727
				2514370F6 (LIVRTUT04)	2303	2825
				1502510F6 (BRAITUT07)	1	439
				70271734V1	183	768
				70273052V1	431	930
				70271651V1	891	1453
31	2817822CB1	1718	1-71, 609-914	2817822F6 (BRSTNOT14)	981	1538
				70272460V1	1094	1718

Table 4 (cont.)

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragments	Sequence Fragments	5' Position	3' Position
32	4009329CB1	2000	1-962	6466193H1 (PLACFEB01)	1	640
				6780428J1 (OVARDIR01)	582	1260
				6307863H1 (NERDTN03)	725	1364
				6781250H1 (OVARDIR01)	972	1639
				7253109J1 (PROSTME05)	1514	1842
				6759035J1 (HEAONOR01)	1515	2000
				5722362H1 (SEMVNOT05)	1	581
33	6618083CB1	2216	1-96, 1201-2216	70789558V1	504	1127
				70787652V1	588	1203
				70791819V1	1050	1650
				70787819V1	1361	1984
				70791126V1	1695	2216
				g2121300.v113.gs_2.nt.edit	1	1995
34	7472002CB1	1995	1-862, 1766-1995	1812692F6 (PROSTUT12)	564	984
				5425924F6 (PROSTMT07)	1	488
				g2525933	823	988
				5000833F6 (PROSTUT21)	283	804
				224000R6 (PANCNOT01)	2435	3087
				6825934J1 (SINTNOR01)	1	515
				7062063H1 (PENITMN02)	2683	3179
35	1812692CB1	988	1-147, 244-570	4491105H1 (BRAMDT02)	2167	2861
				1698347F6 (BLADTUT05)	1762	2333
				70053653D1	1392	1870
				1807402F6 (SINTNOR13)	476	1019
				70055908D1	1317	1837
				7170824H2 (BRSTTMC01)	719	1373
				6555265H1 (BRAFNOR02)	1859	2372
36	3232992CB1	3179	2106-2665, 1-1646	g1444660	992	1464
				g1009986	768	1254
				027195T6 (SPLNFET01)	1674	1986
				g1505781	552	1252
				3358383T6 (PROSTUT16)	1341	1691
				6221856U1	585	1252
				6221857U1	1	727
37	3358383CB1	1986	1465-1986, 1340-1371			

Table 4 (cont.)

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragments	Sequence Fragments	5' Position	3' Position
38	4250091CB1	3294	1-920, 1991-2034, 2488-2760, 1365-1630	g715570	2893	3294
				70759966V1	1864	2477
				4250091F6 (BRADDIR01)	1	532
				5715843H1 (PANCNOT16)	2539	3235
				70789723V1	444	1032
				966456R6 (BRSTNOT05)	2862	3293
				70759467V1	1199	1842
				70788682V1	604	1302
				7056848H1 (BRALNON02)	2373	2989
				858645R1 (BRAITUT03)	1940	2507
39	70064803CB1	2043	1-22, 544-1285	70761829V1	1333	1947
				2758549R6 (THP1AZS08)	1540	2043
				6810024J1 (SKIRNOR01)	576	1280
				1676182T6 (BLADNOT05)	1361	2019
				2109762R6 (BRAITUT03)	1181	1797
				70503885V1	501	1183
				7177480H2 (BRAXDIC01)	1	534
				70450108V1	509	1081
				70451575V1	1072	1730
				1468307F6 (PANCUTUT02)	1	524
40	70356768CB1	1915	1241-1263, 853-897, 1-143, 1450-1532, 668-820	70451567V1	368	1078
				70449058V1	1384	1915
				70449392V1	1060	1720
				6776218J1 (OVARDIR01)	1078	1809
				3024042H1 (PROSDIN01)	690	1040
				6292787H1 (BMARUNA01)	939	1290
				6776218H1 (OVARDIR01)	184	944
				g5686663.v113.gs_16.nt	1	1311
				2613664F6 (ESOGTUT02)	655	1177
				SXBC01035V1	75	573
41	5674114CB1	1809	1-402	2863343F6 (KIDNNOT20)	1	515
				2614317T6 (GBLANOT01)	1126	1730
				SCSA01493V1	548	724
				3323244T6 (PTHYNOT03)	960	1627
				SCIA02891V1	368	1147
				SCIA04658V1	1	641
42	1254635CB1	1730	1-106, 1711-1730, 567-635, 696-900	1-106,		
				1711-1730,		
				567-635,		
				696-900		
43	1670595CB1	1147	746-980, 1-696			

Table 4 (cont.)

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragments	Sequence Fragments	5' Position	3' Position
44	1859560CB1	2745	1-820, 865-2059	6195927H1 (PITUNON01)	2367	2745
				824186R1 (PROSNOT06)	1159	1718
				1399644F6 (BRAITUT08)	503	973
				6812696J1 (ADRETUR01)	30	732
				7255024H1 (FIBRTXC01)	814	1364
				2127239R7 (KIDNNOT05)	1732	2176
				5990868H1 (FTUBTUT02)	1493	1795
				6826978H1 (SINTNOR01)	1	479
				6850265H1 (BRAIFEN08)	2056	2740
				4677711H1 (NOSEDIT02)	1107	1381
				1859560T6 (PROSNOT18)	2247	2745
				4320381H1 (BRADDIT02)	1925	2202
				7175713H1 (BRSTTMC01)	178	811
				3217236H1 (TESTNOT07)	530	812
45	5530164CB1	3204	1241-2064, 1-548, 572-1097	2552002H1 (LUNGUT06)	1	242
				70039789V1	2452	3175
				70090155V1	1781	2492
				6830248J1 (SINTNOR01)	542	1211
				6729730H1 (COLITUT02)	1214	1889
				2850920F6 (BRSTTUT13)	929	1416
				6125934H1 (BRAHNON05)	2505	3204
				6059181H1 (BRAENOT04)	1923	2496
				1956752F6 (CONNNOT01)	1452	1893
				6455739H1 (COLNDIC01)	528	1219
				70077060U1	1900	2535
				7126228H1 (COLNDIY01)	1	580
				2468105F6 (THYRNOT08)	827	1463
				70079483U1	2145	2763
46	139115CB1	2763	1-770, 1345-1389, 1455-1683	70122236V1	1424	1971
				70122363V1	1451	1976
				351595R1 (LVENNOT01)	2066	2566
				70480521V1	1027	1639
				4335403F6 (KIDCTMT01)	1	516
				70466195V1	559	1157
				70466476V1	494	1102
47	1702940CB1	1639	1-246, 1536-1639			

Table 4 (cont.)

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragments	Sequence Fragments	5' Position	3' Position
48	1703342CB1	1600	1-812	3348562H1 (BRAITUT24)	1	282
				285125R1 (EOSIHET02)	1041	1598
				7071066H1 (BRAUTDR02)	250	854
				6494627H1 (BONRNOT01)	1220	1600
49	1727529CB1	2380	1-569, 1228-1654	6879086J1 (LNODNOR03)	360	1094
				957891H1 (KIDNNOT05)	2085	2380
				60211961U1	691	1237
				6800135J1 (COLENOR03)	1250	1983
50	2289333CB1	3038	1-611, 2497-2524	6798918J1 (COLENOR03)	1693	2284
				60211964U1	262	807
				6798894H1 (COLENOR03)	1089	1774
				3249035F6 (SEMYNOT03)	1	626
51	2720354CB1	2608	1-2058	3566495H1 (BRONNOT02)	1984	2298
				2552315T6 (LUNGUT06)	1322	1862
				9872898	848	1328
				1435329F1 (PANCNOT08)	2197	2725
51	2720354CB1	2608	1-2058	3553901H1 (SYNONOT01)	2570	2865
				2508452H1 (CONUTUT01)	1	114
				2771704H1 (COLANOT02)	1815	2078
				6999443H1 (HEALDIR01)	2	553
51	2720354CB1	2608	1-2058	91665184	2594	3038
				2289333R6 (BRAINON01)	1254	1708
				91156003	2524	3032
				5597992H1 (UTRENON03)	1029	1286
51	2720354CB1	2608	1-2058	95545742	612	1066
				5836345H1 (BRAIDIT05)	2624	2880
				4220788F6 (PANCNOT07)	613	958
				1994713T6 (BRSTTUT03)	1949	2451
51	2720354CB1	2608	1-2058	2040880R6 (HIPONON02)	463	821
				2720354F6 (LUNGUT10)	490	1046
				6942433H1 (FTUBTUR01)	885	1437
				6121303H1 (BRAHNON05)	1630	2340
51	2720354CB1	2608	1-2058	6558224H1 (BRAFNON02)	1713	2406
				6940932H1 (FTUBTUR01)	1	465
				91927466	325	872
				6826181J1 (SINTNOR01)	1062	1666
51	2720354CB1	2608	1-2058	6197805H1 (PITUNON01)	2154	2608

Table 4 (cont.)

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragments	Sequence Fragments	5' Position	3' Position
52	3038193CB1	3804	3392-3457, 1169-1264, 1-829, 2271-2483, 1319-1363	044564H1 (TBLYNOT01) 901446R6 (BRSTTUT03) 94088232 1428831H1 (SINTBST01) 2741328T6 (BRSTTUT14) 4970206H1 (KIDEUNC10) 2768967H1 (COLANOT02) 5688762F6 (BRAIUNT01) 3038193F6 (BRSTNOT16) 6477440H1 (PROSTMC01) 70809191V1 3154867H1 (TLYMTXT02) 2257401R6 (OVARTUT01) 94268882 2257401T6 (OVARTUT01) 2237852F6 (PANCFTUT02) 3460979F6 (293TFLT01) 7161336H1 (PLACNOR01) 6800921J1 (COLENOR03) 7057496H1 (BRALNON02) GNN.g6554406_006	2311 1733 3459 473 3235 1029 772 36 1284 1843 2411 1 2660 1421 2896 519 1000 582 1 1206 1	2562 2282 3804 661 3804 1303 1020 621 1710 2486 2832 272 3167 1815 3520 945 1500 1193 557 1894 1668
53	3460979CB1	1894	1-36 1746-1894			
54	7472200CB1	1668	1-1668			

Table 5

Polynucleotide SEQ ID NO:	Incyte Project ID	Representative Library
28	1416107CB1	UTREDIT07
29	1682513CB1	SPLNNT011
30	2446438CB1	MCLDTXN03
31	2817822CB1	BRAITUT07
32	4009329CB1	OVARDIN02
33	6618083CB1	HELAUNT01
35	1812692CB1	PROSTUT12
36	3232992CB1	PANCN0T01
37	3358383CB1	PROSTUT16
38	4250091CB1	BRAITUT03
39	70064803CB1	THP1AZS08
40	70356768CB1	HNT2AGT01
41	5674114CB1	OVARDIR01
42	1254635CB1	LUNGFET03
43	1670595CB1	BRAITUT24
44	1859560CB1	NGANN0T01
45	5530164CB1	BRAYDIN03
46	139115CB1	SINTNOT18
47	1702940CB1	BRAVXT04
48	1703342CB1	EOSIHET02
49	1727529CB1	PROSNOT18
50	2289333CB1	LUNGITUT06
51	2720354CB1	PROSTUS23
52	3038193CB1	LIVRN0N08
53	3460979CB1	COLENOR03

Table 6

Library	Vector	Library Description
UTREDIT07	pINCY	Library was constructed using RNA isolated from diseased endometrial tissue removed from a female during endometrial biopsy. Pathology indicated in phase endometrium with missing beta 3, Type II defects.
SPLNNOT11	pINCY	Library was constructed using RNA isolated from diseased spleen tissue removed from a 14-year-old Asian male during a total splenectomy. Pathology indicated changes consistent with idiopathic thrombocytopenic purpura. The patient presented with bruising. Patient medications included Vincristine.
MCLDTXN03	pINCY	Library was constructed from a pool of two dendritic cell libraries. Starting libraries were constructed using RNA isolated from untreated and treated derived dendritic cells from umbilical cord blood CD34+ precursor cells removed from a male. The cells were derived with granulocyte/macrophage colony stimulating factor (GM-CSF), tumor necrosis factor alpha (TNF alpha), and stem cell factor (SCF). The libraries were normalized under conditions adapted from Soares et al. (1994) Proc. Natl. Acad. Sci. USA 91:9228 and Bonaldo et al. (1996) Genome Res. 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
BRAITUT07	pINCY	Library was constructed using RNA isolated from left frontal lobe tumor tissue removed from the brain of a 32-year-old Caucasian male during excision of a cerebral meningeal lesion. Pathology indicated low grade desmoplastic neuronal neoplasm. The patient presented with nausea, vomiting, and headache. Patient history included alcohol, tobacco use, and marijuana use twice a week for six years. Family history included atherosclerotic coronary artery disease in the grandparent(s).
OVARDIN02	pINCY	Library was constructed from an ovarian tissue library. Starting RNA was made from diseased ovarian tissue removed from a 39-year-old Caucasian female during total abdominal hysterectomy, bilateral salpingo-oophorectomy, dilation and curettage, partial colectomy, incidental appendectomy, and temporary colostomy. Pathology indicated the right and left adnexa, mesentery and muscularis propria of the sigmoid colon were extensively involved by endometriosis. Endometriosis also involved the anterior and posterior serosal surfaces of the uterus and the cul-de-sac. The endometrium was proliferative. Pathology for the associated tumor tissue indicated multiple (3 intramural, 1 subserosal) leiomyomata. The patient presented with abdominal pain and infertility. Patient history included scoliosis. Previous surgeries included laparoscopic cholecystectomy and exploratory laparotomy. Patient medications included Megace, Danazol, and Lupron. Family history included hyperlipidemia in the mother, benign hypertension, hyperlipidemia, atherosclerotic coronary artery disease, coronary artery bypass graft, depressive disorder, brain cancer, and type II diabetes. The library was normalized under conditions adapted from Soares et al. (1994) Proc. Natl. Acad. Sci. USA 91:9228 and Bonaldo et al. (1996) Genome Res. 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
HELAUNT01	pINCY	Library was constructed from RNA isolated from an untreated HeLa cell line, derived from cervical adenocarcinoma removed from a 31-year-old Black female.

Table 6 (cont.)

Library	Vector	Library Description
BRAITUT03	PSPORT1	Library was constructed using RNA isolated from brain tumor tissue removed from the left frontal lobe of a 17-year-old Caucasian female during excision of a cerebral meningeal lesion. Pathology indicated a grade 4 fibrillary giant and small-cell astrocytoma. Family history included benign hypertension and cerebrovascular disease.
HNT2AGT01	PBLUESCRIPT	Library was constructed at Stratagene (STR937233), using RNA isolated from the hNT2 cell line derived from a human teratocarcinoma that exhibited properties characteristic of a committed neuronal precursor. Cells were treated with retinoic acid for 5 weeks and with mitotic inhibitors for two weeks and allowed to mature for an additional 4 weeks in conditioned medium.
OVARDIR01	pcDNA2.1	Library was constructed using RNA isolated from right ovary tissue removed from a 45-year-old Caucasian female during total abdominal hysterectomy, bilateral salpingo-oophorectomy, vaginal suspension and fixation, and incidental appendectomy. Pathology indicated stromal hyperthecosis of the right and left ovaries. Pathology for the matched tumor tissue indicated a dermoid cyst (benign cystic teratoma) in the left ovary. Multiple (3) intramural leiomyomata were identified. The cervix showed squamous metaplasia. Patient history included metrorrhagia, female stress incontinence, alopecia, depressive disorder, pneumonia, normal delivery, and deficiency anemia. Family history included benign hypertension, atherosclerotic coronary artery disease, hyperlipidemia, and primary tuberculous complex.
PANCN0T01	PBLUESCRIPT	Library was constructed using RNA isolated from the pancreatic tissue of a 29-year-old Caucasian male who died from head trauma.
PROSTUT12	pINCY	Library was constructed using RNA isolated from prostate tumor tissue removed from a 65-year-old Caucasian male during a radical prostatectomy. Pathology indicated an adenocarcinoma (Gleason grade 2+2). Adenofibromatous hyperplasia was also present. The patient presented with elevated prostate specific antigen (PSA).
PROSTUT16	pINCY	Library was constructed using RNA isolated from prostate tumor tissue removed from a 55-year-old Caucasian male. Pathology indicated adenocarcinoma, Gleason grade 5+4. Adenofibromatous hyperplasia was also present. The patient presented with elevated prostate specific antigen (PSA). Patient history included calculus of the kidney. Family history included lung cancer and breast cancer.
THP1AZS08	PSPORT1	This subtracted THP-1 promonocyte cell line library was constructed using 5.76 x 1e6 clones from a 5-aza-2'-deoxycytidine (AZ) treated THP-1 cell library. Starting RNA was made from THP-1 promonocyte cells treated for three days with 0.8 micromolar AZ. The hybridization probe for subtraction was derived from a similarly constructed library, made from RNA isolated from untreated THP-1 cells. 5.76 million clones from the AZ-treated THP-1 cell library were then subjected to two rounds of subtractive hybridization with 5 million clones from the untreated THP-1 cell library. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991) 19:1954, and Donaldo et al., Genome Research (1996) 6:791. THP-1 (ATCC TIB 202) is a human promonocyte line derived from peripheral blood of a 1-year-old Caucasian male with acute monocytic leukemia (ref: Int. J. Cancer 26 (1980):171).

Table 6 (cont.)

Library	Vector	Library Description
BRAITUT24	pINCY	Library was constructed using RNA isolated from right frontal brain tumor tissue removed from a 50-year-old Caucasian male during a cerebral meninges lesion excision. Pathology indicated meningioma. Family history included colon cancer and cerebrovascular disease.
BRAYDIN03	pINCY	This normalized brain tissue library was constructed from 6.7 million independent clones from a brain tissue library. Starting RNA was made from RNA isolated from diseased hypothalamus tissue removed from a 57-year-old Caucasian male who died from a cerebrovascular accident. Patient history included Huntington's disease and emphysema. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used. The library was linearized and recircularized to select for insert containing clones.
LUNGFET03	pINCY	Library was constructed using RNA isolated from lung tissue removed from a Caucasian female fetus, who died at 20 weeks' gestation.
NGANNOT01	PSPORT1	Library was constructed using RNA isolated from tumorous neuroganglion tissue removed from a 9-year-old Caucasian male during a soft tissue excision of the chest wall. Pathology indicated a ganglioneuroma. Family history included asthma.
BRAVXT04	PSPORT1	Library was constructed using RNA isolated from separate populations of human astrocytes stimulated for 4 to 6 hours with a combination of cytokines including IL-1. The RNA was pooled for polyA RNA isolation and library construction.
EOSIHET02	PBLUESCRIPT	Library was constructed using RNA isolated from peripheral blood cells adhered from a 48-year-old Caucasian male. Patient history included hypereosinophilia. The cell population was determined to be greater than 77% eosinophils by Wright's staining.
LIVRNON08	pINCY	This normalized liver tissue library was constructed from 5.7 million independent clones from a pooled liver tissue library. Starting RNA was isolated from pooled liver tissue removed from a 4-year-old Hispanic male who died from anoxia and a 16 week female fetus who died after 16-weeks gestation from anencephaly. Serologies were positive for cytomegalovirus in the 4-year-old. Patient history included asthma in the 4-year-old. Family history included taking daily prenatal vitamins and mitral valve prolapse in the mother of the fetus. The library was normalized in 2 rounds using conditions adapted from Soares et al. Proc. Natl. Acad. Sci. USA (1994) 91:9228 and Bonaldo et al. (1996) Genome Research 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.

Table 6 (cont.)

Library	Vector	Library Description
LUNGTUT06	pINCY	Library was constructed using RNA isolated from apical lung tumor tissue removed from an 80-year-old Caucasian female during a segmental lung resection. Pathology indicated a metastatic granulosa cell tumor. Patient history included pelvic soft tissue tumor and chemotherapy for one year. Family history included tuberculosis, lung cancer, and atherosclerotic coronary artery disease.
PROSNOT18	pINCY	Library was constructed using RNA isolated from diseased prostate tissue removed from a 58-year-old Caucasian male during a radical cystectomy, radical prostatectomy, and gastrectomy. Pathology indicated adenofibromatous hyperplasia; this tissue was associated with a grade 3 transitional cell carcinoma. Patient history included angina and emphysema. Family history included acute myocardial infarction, atherosclerotic coronary artery disease, and type II diabetes.
PROSTUS23	pINCY	This subtracted prostate tumor library was constructed using 1 million clones from a pooled prostate tumor library that was subjected to 2 rounds of subtractive hybridization with 1 million clones from a pooled prostate tissue library. The starting library for subtraction was constructed by pooling equal numbers of clones from 4 prostate tumor libraries using mRNA isolated from prostate tumor removed from Caucasian males at ages 58 (A), 61 (B), 66 (C), and 68 (D) during prostatectomy with lymph node excision. Pathology indicated adenocarcinoma in all donors. History included elevated PSA, induration and tobacco abuse in donor A; elevated PSA, induration, prostate hyperplasia, renal failure, osteoarthritis, renal artery stenosis, benign HTN, thrombocytopenia, hyperlipidemia, tobacco/alcohol and hepatitis C (carrier) in donor B; elevated PSA, induration, and tobacco abuse in donor C; and elevated PSA, induration, hypercholesterolemia, and kidney calculus in donor D. The hybridization probe for subtraction was constructed by pooling equal numbers of cDNA clones from 3 prostate tissue libraries derived from prostate tissue, prostate epithelial cells, and fibroblasts from prostate stroma from 3 different donors. Subtractive hybridization conditions were based on the methodologies of Swaroop et al. (1991) Nucleic Acids Res. 19:1954 and Bonaldo et al. Genome Research (1996) 6:791.
SINTNOT18	pINCY	Library was constructed using RNA isolated from small intestine tissue obtained from a 59-year-old male.
COLENOR03	PCDNA2.1	Library was constructed using RNA isolated from colon epithelium tissue removed from a 13-year-old Caucasian female who died from a motor vehicle accident.

Table 7

Program	Description	Reference	Parameter Threshold
ABI FACTURA	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less
FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises at least five functions: fasta, tfasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad. Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489.	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Probability value= 1.0E-3 or less
HMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, E.L.L. et al. (1998) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350.	PFAM hits: Probability value= 1.0E-3 or less Signal peptide hits: Score= 0 or greater

Table 7 (cont.)

Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality score \geq GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	

What is claimed is:

1. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of:

- 5 a) an amino acid sequence selected from the group consisting of SEQ ID NO:1-27,
 b) a naturally occurring amino acid sequence having at least 90% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO:1-27,
 c) a biologically active fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27, and
10 d) an immunogenic fragment of an amino acid sequence selected from the group consisting of SEQ ID NO:1-27.

2. An isolated polypeptide of claim 1 selected from the group consisting of SEQ ID NO:1-27.

15

3. An isolated polynucleotide encoding a polypeptide of claim 1.

4. An isolated polynucleotide encoding a polypeptide of claim 2.

20 5. An isolated polynucleotide of claim 4 selected from the group consisting of SEQ ID NO:28-54.

6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.

25

7. A cell transformed with a recombinant polynucleotide of claim 6.

8. A transgenic organism comprising a recombinant polynucleotide of claim 6.

30 9. A method for producing a polypeptide of claim 1, the method comprising:

 a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and

35 b) recovering the polypeptide so expressed.

10. An isolated antibody which specifically binds to a polypeptide of claim 1.

11. An isolated polynucleotide comprising a polynucleotide sequence selected from the group consisting of:

- 5 a) a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54,
 b) a naturally occurring polynucleotide sequence having at least 90% sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:28-54,
 c) a polynucleotide sequence complementary to a),
 d) a polynucleotide sequence complementary to b), and
10 e) an RNA equivalent of a)-d).

12. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 11.

15 13. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:

- a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization
20 complex is formed between said probe and said target polynucleotide or fragments thereof, and
 b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.

25 14. A method of claim 13, wherein the probe comprises at least 60 contiguous nucleotides.

 15. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:

- a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
30 b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

 16. A composition comprising an effective amount of a polypeptide of claim 1 and a pharmaceutically acceptable excipient.

35

17. A composition of claim 16, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-27.

18. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition of claim 16.

19. A method for screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:

- a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- b) detecting agonist activity in the sample.

20. A composition comprising an agonist compound identified by a method of claim 19 and a pharmaceutically acceptable excipient.

21. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 20.

22. A method for screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:

- a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- b) detecting antagonist activity in the sample.

23. A composition comprising an antagonist compound identified by a method of claim 22 and a pharmaceutically acceptable excipient.

24. A method for treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 23.

25. A method of screening for a compound that specifically binds to the polypeptide of claim 1, said method comprising the steps of:

- a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
- b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a

compound that specifically binds to the polypeptide of claim 1.

26. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, said method comprising:

- 5 a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
- b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
- c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in
- 10 the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.

27. A method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method

15 comprising:

- a) exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
- b) detecting altered expression of the target polynucleotide, and
- c) comparing the expression of the target polynucleotide in the presence of varying amounts of
- 20 the compound and in the absence of the compound.

28. A method for assessing toxicity of a test compound, said method comprising:

- a) treating a biological sample containing nucleic acids with the test compound;
- b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at
- 25 least 20 contiguous nucleotides of a polynucleotide of claim 11 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 11 or fragment thereof;
- c) quantifying the amount of hybridization complex; and
- 30 d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

<110> INCYTE GENOMICS, INC.
 BAUGHN, Mariah R.
 BURFORD, Neil
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 LU, Dyung Aina M.
 YANG, Junming
 REDDY, Roopa
 LAL, Preeti
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 AZIMZAI, Yalda
 YUE, Henry
 NGUYEN, Danniel B.
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Lys	Gln	Phe	Leu	Leu	Gln	Val	Pro	Gly	Tyr	Ser	Glu	Asn	Leu	Glu	
				140					145					150	
Leu	Met	Val	Arg	Leu	Ala	Arg	Ala	Glu	Ala	Ile	Thr	Ala	Arg	Lys	
				155					160					165	
Ser	Ser	Val	Leu	Val	Cys	Leu	Val	Glu	Thr	Glu	Glu	Gln	Asp	Ala	
				170					175					180	
Tyr	Tyr	Ser	Glu	Val	Leu	Cys	Phe	Leu	Gln	Asp	Lys	Lys	Met	Phe	
				185					190					195	

Lys	Ser	Val	Val	Lys	Phe	Gly	Pro	Trp	Lys	Ala	Val	Leu	Asp	Asn
				200					205					210
Ser	Asp	Leu	Met	His	Cys	Leu	Glu	Met	Asp	Ile	Lys	Ala	Gln	Gly
				215					220					225
Tyr	Lys	Val	Phe	Ser	Lys	Phe	Tyr	Leu	Thr	Tyr	Pro	Thr	Lys	Arg
				230					235					240
Asn	Glu	Phe	His	Phe	Asn	Ile	Tyr	Ser	Phe	Thr	Phe	Thr	Trp	Trp
				245					250					255

<210> 5

<211> 584

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 4009329CD1

<400> 5

Met	Ala	Gly	Arg	Arg	Leu	Asn	Leu	Arg	Trp	Ala	Leu	Ser	Val	Leu
1				5					10					15
Cys	Val	Leu	Leu	Met	Ala	Glu	Thr	Val	Ser	Gly	Thr	Arg	Gly	Ser
				20					25					30
Ser	Thr	Gly	Ala	His	Ile	Ser	Pro	Gln	Phe	Pro	Ala	Ser	Gly	Val
				35					40					45
Asn	Gln	Thr	Pro	Val	Val	Asp	Cys	Arg	Lys	Val	Cys	Gly	Leu	Asn
				50					55					60
Val	Ser	Asp	Arg	Cys	Asp	Phe	Ile	Arg	Thr	Asn	Pro	Asp	Cys	His
				65					70					75
Ser	Asp	Gly	Gly	Tyr	Leu	Asp	Tyr	Leu	Glu	Gly	Ile	Phe	Cys	His
				80					85					90
Phe	Pro	Pro	Ser	Leu	Leu	Pro	Leu	Ala	Val	Thr	Leu	Tyr	Val	Ser
				95					100					105
Trp	Leu	Leu	Tyr	Leu	Phe	Leu	Ile	Leu	Gly	Val	Thr	Ala	Ala	Lys
				110					115					120
Phe	Phe	Cys	Pro	Asn	Leu	Ser	Ala	Ile	Ser	Thr	Thr	Leu	Lys	Leu
				125					130					135
Ser	His	Asn	Val	Ala	Gly	Val	Thr	Phe	Leu	Ala	Phe	Gly	Asn	Gly
				140					145					150
Ala	Pro	Asp	Ile	Phe	Ser	Ala	Leu	Val	Ala	Phe	Ser	Asp	Pro	His
				155					160					165
Thr	Ala	Gly	Leu	Ala	Leu	Gly	Ala	Leu	Phe	Gly	Ala	Gly	Val	Leu
				170					175					180
Val	Thr	Thr	Val	Val	Ala	Gly	Gly	Ile	Thr	Ile	Leu	His	Pro	Phe
				185					190					195
Met	Ala	Ala	Ser	Arg	Pro	Phe	Phe	Arg	Asp	Ile	Val	Phe	Tyr	Met
				200					205					210
Val	Ala	Val	Phe	Leu	Thr	Phe	Leu	Met	Leu	Phe	Arg	Gly	Arg	Val
				215					220					225
Thr	Leu	Ala	Trp	Ala	Leu	Gly	Tyr	Leu	Gly	Leu	Tyr	Val	Phe	Tyr
				230					235					240
Val	Val	Thr	Val	Ile	Leu	Cys	Thr	Trp	Ile	Tyr	Gln	Arg	Gln	Arg
				245					250					255
Arg	Gly	Ser	Leu	Phe	Cys	Pro	Met	Pro	Val	Thr	Pro	Glu	Ile	Leu
				260					265					270
Ser	Asp	Ser	Glu	Glu	Asp	Arg	Val	Ser	Ser	Asn	Thr	Asn	Ser	Tyr
				275					280					285
Asp	Tyr	Gly	Asp	Glu	Tyr	Arg	Pro	Leu	Phe	Phe	Tyr	Gln	Glu	Thr
				290					295					300
Thr	Ala	Gln	Ile	Leu	Val	Arg	Ala	Leu	Asn	Pro	Leu	Asp	Tyr	Met
				305					310					315
Lys	Trp	Arg	Arg	Lys	Ser	Ala	Tyr	Trp	Lys	Ala	Leu	Lys	Val	Phe
				320					325					330
Lys	Leu	Pro	Val	Glu	Phe	Leu	Leu	Leu	Leu	Thr	Val	Pro	Val	Val
				335					340					345
Asp	Pro	Asp	Lys	Asp	Asp	Gln	Asn	Trp	Lys	Arg	Pro	Leu	Asn	Cys

Leu His Leu Val	350	Ile Ser Pro Leu Val	355	Val Val Leu Thr Leu	360
	365		370		375
Ser Gly Thr Tyr	Gly Val Tyr Glu Ile	Gly Gly Leu Val Pro	Val		
	380		385		390
Trp Val Val Val	Val Ile Ala Gly Thr	Ala Leu Ala Ser Val	Thr		
	395		400		405
Phe Phe Ala Thr	Ser Asp Ser Gln Pro	Pro Arg Leu His Trp	Leu		
	410		415		420
Phe Ala Phe Leu	Gly Phe Leu Thr Ser	Ala Leu Trp Ile Asn	Ala		
	425		430		435
Ala Ala Thr Glu	Val Val Asn Ile Leu	Arg Ser Leu Gly Val	Val		
	440		445		450
Phe Arg Leu Ser	Asn Thr Val Leu Gly	Leu Thr Leu Leu Ala	Trp		
	455		460		465
Gly Asn Ser Ile	Gly Asp Ala Phe Ser	Asp Phe Thr Leu Ala	Arg		
	470		475		480
Gln Gly Tyr Pro	Arg Met Ala Phe Ser	Ala Cys Phe Gly Gly	Ile		
	485		490		495
Ile Phe Asn Ile	Leu Val Gly Val Gly	Leu Gly Cys Leu Leu	Gln		
	500		505		510
Ile Ser Arg Ser	His Thr Glu Val Lys	Leu Glu Pro Asp Gly	Leu		
	515		520		525
Leu Val Trp Val	Leu Ala Gly Ala Leu	Gly Leu Ser Leu Val	Phe		
	530		535		540
Ser Leu Val Ser	Val Pro Leu Gln Cys	Phe Gln Leu Ser Arg	Val		
	545		550		555
Tyr Gly Phe Cys	Leu Leu Leu Phe Tyr	Leu Asn Phe Leu Val	Val		
	560		565		570
Ala Leu Leu Ile	Glu Phe Gly Val Ile	His Leu Lys Ser Met			
	575		580		

<210> 6

<211> 416

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 6618083CD1

<400> 6

Met Lys Leu Ser Lys	Lys Asp Arg Gly Glu	Asp Glu Glu Ser Asp	
1	5	10	15
Ser Ala Lys Lys Lys	Leu Asp Trp Ser Cys	Ser Leu Leu Val Ala	
	20	25	30
Ser Leu Ala Gly Ala	Phe Gly Ser Ser Phe	Leu Tyr Gly Tyr Asn	
	35	40	45
Leu Ser Val Val Asn	Ala Pro Thr Pro Tyr	Ile Lys Ala Phe Tyr	
	50	55	60
Asn Glu Ser Trp Glu	Arg Arg His Gly Arg	Pro Ile Asp Pro Asp	
	65	70	75
Thr Leu Thr Leu Leu	Trp Ser Val Thr Val	Ser Ile Phe Ala Ile	
	80	85	90
Gly Gly Leu Val Gly	Thr Leu Ile Val Lys	Met Ile Gly Lys Val	
	95	100	105
Leu Gly Arg Lys His	Thr Leu Leu Ala Asn	Asn Gly Phe Ala Ile	
	110	115	120
Ser Ala Ala Leu Leu	Met Ala Cys Ser Leu	Gln Ala Gly Ala Phe	
	125	130	135
Glu Met Leu Ile Val	Gly Arg Phe Ile Met	Gly Ile Asp Gly Gly	
	140	145	150
Val Ala Leu Ser Val	Leu Pro Met Tyr Leu	Ser Glu Ile Ser Pro	
	155	160	165
Lys Glu Ile Arg Gly	Ser Leu Gly Gln Val	Thr Ala Ile Phe Ile	
	170	175	180
Cys Ile Gly Val Phe	Thr Gly Gln Leu Leu	Gly Leu Pro Glu Leu	

Leu Gly Lys Glu Ser	Thr Trp Pro Tyr	Leu Phe Gly Val Ile Val	185	190	195
Val Pro Ala Val Val	Gln Leu Leu Ser	Leu Pro Phe Leu Pro Asp	200	205	210
Ser Pro Arg Tyr Leu	Leu Leu Glu Lys	His Asn Glu Ala Arg Ala	215	220	225
Val Lys Ala Phe Gln	Thr Phe Leu Gly	Lys Ala Asp Val Ser Gln	230	235	240
Glu Val Glu Glu Val	Leu Ala Glu Ser	His Val Gln Arg Ser Ile	245	250	255
Arg Leu Val Ser Val	Leu Glu Leu Leu	Arg Ala Pro Tyr Val Arg	260	265	270
Trp Gln Val Val Thr	Val Ile Val Thr	Met Ala Cys Tyr Gln Leu	275	280	285
Cys Gly Leu Asn Ala	Ile Trp Phe Tyr	Thr Asn Ser Ile Phe Gly	290	295	300
Lys Ala Gly Ile Pro	Leu Ala Lys Ile	Pro Tyr Val Thr Leu Ser	305	310	315
Thr Gly Gly Ile Glu	Thr Leu Ala Ala	Val Phe Ser Gly Leu Val	320	325	330
Ile Glu His Leu Gly	Arg Arg Pro Leu	Leu Ile Gly Gly Phe Gly	335	340	345
Leu Met Gly Leu Phe	Phe Gly Thr Leu	Thr Ile Thr Leu Thr Leu	350	355	360
Gln Asp His Ala Pro	Trp Val Pro Tyr	Leu Ser Ile Val Gly Ile	365	370	375
Leu Ala Ile Ile Ala	Ser Phe Cys Ser	Gly Pro Ala Val Phe Pro	380	385	390
Glu Glu Thr Val Asn	Val Ser Ile Val	Ser Glu	395	400	405
			410	415	

<210> 7

<211> 664

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 7472002CD1

<400> 7

Met Thr Glu Lys Thr	Asn Gly Val Lys Ser	Ser Pro Ala Asn Asn	1	5	10	15
His Asn His His Ala	Pro Pro Ala Ile Lys	Ala Asn Gly Lys Asp	20	25	30	35
Asp His Arg Thr Ser	Ser Arg Pro His Ser	Ala Ala Asp Asp Asp	40	45	50	55
Thr Ser Ser Glu Leu	Gln Arg Leu Ala Asp	Val Asp Ala Pro Gln	60	65	70	75
Gln Gly Arg Ser Gly	Phe Arg Arg Ile Val	Arg Leu Val Gly Ile	80	85	90	95
Ile Arg Glu Trp Ala	Asn Lys Asn Phe Arg	Glu Glu Glu Pro Arg	100	105	110	115
Pro Asp Ser Phe Leu	Glu Arg Phe Arg Gly	Pro Glu Leu Gln Thr	120	125	130	135
Val Thr Thr Gln Glu	Gly Asp Gly Lys Gly	Asp Lys Asp Gly Glu	140	145	150	155
Asp Lys Gly Thr Lys	Lys Lys Phe Glu Leu	Phe Val Leu Asp Pro	160	165	170	175
Ala Gly Asp Trp Tyr	Tyr Cys Trp Leu Phe	Val Ile Ala Met Pro	180	185	190	195
Val Leu Tyr Asn Trp	Cys Leu Leu Val Ala	Arg Ala Cys Phe Ser	200	205	210	215
Asp Leu Gln Lys Gly	Tyr Tyr Leu Val Trp	Leu Val Leu Asp Tyr	220	225	230	235
Val Ser Asp Val Val	Tyr Ile Ala Asp Leu	Phe Ile Arg Leu Arg	240	245	250	255

Thr Gly Phe Leu	185	Gln Gly Leu Leu	190	Lys Asp Thr Lys	195
	200		205		210
Leu Arg Asp Asn	215	Ile His Thr Leu	220	Phe Lys Leu Asp	225
	230		235		240
Ala Ser Ile Ile	245	Thr Asp Leu Ile	250	Phe Ala Val Asp	255
	260		265		270
His Ser Pro Glu	275	Val Arg Phe Asn Arg	280	Leu Leu His Phe Ala	285
	290		295		300
Met Phe Glu Phe	305	Phe Asp Arg Thr Glu	310	Thr Arg Thr Asn Tyr	315
	320		325		330
Asn Ile Phe Arg	335	Ile Ser Asn Leu Val	340	Tyr Ile Leu Val	345
	350		355		360
Ile His Trp Asn	365	Ala Cys Ile Tyr Tyr	370	Ala Ile Ser Lys Ser	375
	380		385		390
Gly Phe Gly Val	395	Thr Trp Val Tyr	400	Pro Asn Ile Thr Asp	405
	410		415		420
Glu Tyr Gly Tyr	425	Leu Ala Arg Glu Tyr	430	Ile Tyr Cys Leu Tyr	435
	440		445		450
Ser Thr Leu Thr	455	Leu Thr Thr Ile Gly	460	Glu Thr Pro Pro Pro	465
	470		475		480
Lys Asp Glu Glu	485	Tyr Leu Phe Val Ile	490	Phe Asp Phe Leu Ile	495
	500		505		510
Val Leu Ile Phe	515	Ala Thr Ile Val Gly	520	Asn Val Gly Ser Met	525
	530		535		540
Ser Asn Met Asn	545	Ala Thr Arg Ala Glu	550	Phe Gln Ala Lys Ile	555
	560		565		570
Ala Val Lys His	575	Tyr Met Gln Phe Arg	580	Lys Val Ser Lys Gly	585
	590		595		600
Glu Ala Lys Val	605	Ile Arg Trp Phe Asp	610	Tyr Leu Trp Thr Asn	615
	620		625		630
Lys Thr Val Asp	635	Glu Arg Glu Ile Leu	640	Lys Asn Leu Pro Ala	645
	650		655		660
Leu Arg Ala Glu		Ile Ala Ile Asn Val		His Leu Ser Thr Leu	
Lys Val Arg Ile		Phe His Asp Cys Glu		Ala Gly Leu Leu Val	
Leu Val Leu Lys		Leu Arg Pro Gln Val		Phe Ser Pro Gly Asp	
Ile Cys Arg Lys		Gly Asp Ile Gly Lys		Glu Met Tyr Ile Ile	
Glu Gly Lys Leu		Ala Val Val Ala Asp		Asp Gly Val Thr Gln	
Ala Leu Leu Ser		Ala Gly Ser Cys Phe		Gly Glu Ile Ser Ile	
Asn Ile Lys Gly		Ser Lys Met Gly Asn		Arg Arg Thr Ala Asn	
Arg Ser Leu Gly		Tyr Ser Asp Leu Phe		Cys Leu Ser Lys Asp	
Leu Met Glu Ala		Val Thr Glu Tyr Pro		Asp Ala Lys Lys Val	
Glu Glu Arg Gly		Arg Glu Ile Leu Met		Lys Glu Gly Leu Leu	
Glu Asn Glu Val		Ala Thr Ser Met Glu		Val Asp Val Gln Glu	
Leu Gly Gln Leu		Glu Thr Asn Met Glu		Thr Leu Tyr Thr Arg	
Gly Arg Leu Leu		Ala Glu Tyr Thr Gly		Ala Gln Gln Lys Leu	
Gln Arg Ile Thr		Val Leu Glu Thr Lys		Met Lys Gln Asn Asn	
Asp Asp Tyr Leu		Ser Asp Gly Met Asn		Ser Pro Glu Leu Ala	
Ala Asp Glu Pro					

<210> 8

<211> 242
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 1812692CD1

<400> 8
 Met Ser Phe Arg Ala Ala Arg Leu Ser Met Arg Asn Arg Arg Asn
 1 5 10 15
 Asp Thr Leu Asp Ser Thr Arg Thr Leu Tyr Ser Ser Ala Ser Arg
 20 25 30
 Ser Thr Asp Leu Ser Tyr Ser Glu Ser Asp Leu Val Asn Phe Ile
 35 40 45
 Gln Ala Asn Phe Lys Lys Arg Glu Cys Val Phe Phe Thr Lys Asp
 50 55 60
 Ser Lys Ala Thr Glu Asn Val Cys Lys Cys Gly Tyr Ala Gln Ser
 65 70 75
 Gln His Met Glu Gly Thr Gln Ile Asn Gln Ser Glu Lys Trp Asn
 80 85 90
 Tyr Lys Lys His Thr Lys Glu Phe Pro Thr Asp Ala Phe Gly Asp
 95 100 105
 Ile Gln Phe Glu Thr Leu Gly Lys Lys Gly Lys Tyr Ile Arg Leu
 110 115 120
 Ser Cys Asp Thr Asp Ala Glu Ile Leu Tyr Glu Leu Leu Thr Gln
 125 130 135
 His Trp His Leu Lys Thr Pro Asn Leu Val Ile Ser Val Thr Gly
 140 145 150
 Gly Ala Lys Asn Phe Ala Leu Lys Pro Arg Met Arg Lys Ile Phe
 155 160 165
 Ser Arg Leu Ile Tyr Ile Ala Gln Ser Lys Gly Ala Trp Ile Leu
 170 175 180
 Thr Gly Gly Thr His Tyr Gly Leu Met Lys Tyr Ile Gly Glu Val
 185 190 195
 Val Arg Asp Asn Thr Ile Ser Arg Ser Ser Glu Glu Asn Ile Val
 200 205 210
 Ala Ile Gly Ile Ala Ala Trp Gly Met Val Ser Asn Arg Asp Thr
 215 220 225
 Leu Ile Arg Asn Cys Asp Ala Glu Val Pro Val Gly Gln Glu Glu
 230 235 240
 Val Cys

<210> 9
 <211> 398
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 3232992CD1

<400> 9
 Met Val Ala Ala Pro Ile Phe Gly Tyr Leu Gly Asp Arg Phe Asn
 1 5 10 15
 Arg Lys Val Ile Leu Ser Cys Gly Ile Phe Phe Trp Ser Ala Val
 20 25 30
 Thr Phe Ser Ser Ser Phe Ile Pro Gln Gln Tyr Phe Trp Leu Leu
 35 40 45
 Val Leu Ser Arg Gly Leu Val Gly Ile Gly Glu Ala Ser Tyr Ser
 50 55 60
 Thr Ile Ala Pro Thr Ile Ile Gly Asp Leu Phe Thr Lys Asn Thr
 65 70 75
 Arg Thr Leu Met Leu Ser Val Phe Tyr Phe Ala Ile Pro Leu Gly
 80 85 90
 Ser Gly Leu Gly Tyr Ile Thr Gly Ser Ser Val Lys Gln Ala Ala

	95		100		105
Gly Asp Trp His	Trp Ala Leu Arg Val	Ser Pro Val Leu Gly	Met		
	110		115		
Ile Thr Gly Thr	Leu Ile Leu Ile Leu	Val Pro Ala Thr Lys	Arg		
	125		130		
Gly His Ala Asp	Gln Leu Gly Asp Gln	Leu Lys Ala Arg Thr	Ser		
	140		145		
Trp Leu Arg Asp	Met Lys Ala Leu Ile	Arg Asn Arg Ser Tyr	Val		
	155		160		
Phe Ser Ser Leu	Ala Thr Ser Ala Val	Ser Phe Ala Thr Gly	Ala		
	170		175		
Leu Gly Met Trp	Ile Pro Leu Tyr Leu	His Arg Ala Gln Val	Val		
	185		190		
Gln Lys Thr Ala	Glu Thr Cys Asn Ser	Pro Pro Cys Gly Ala	Lys		
	200		205		
Asp Ser Leu Ile	Phe Gly Ala Ile Thr	Cys Phe Thr Gly Phe	Leu		
	215		220		
Gly Val Val Thr	Gly Ala Gly Ala Thr	Arg Trp Cys Arg Leu	Lys		
	230		235		
Thr Gln Arg Ala	Asp Pro Leu Val Cys	Ala Val Gly Met Leu	Gly		
	245		250		
Ser Ala Ile Phe	Ile Cys Leu Ile Phe	Val Ala Ala Lys Ser	Ser		
	260		265		
Ile Val Gly Ala	Tyr Ile Cys Ile Phe	Val Gly Glu Thr Leu	Leu		
	275		280		
Phe Ser Asn Trp	Ala Ile Thr Ala Asp	Ile Leu Met Tyr Val	Val		
	290		295		
Ile Pro Thr Arg	Arg Ala Thr Ala Val	Ala Leu Gln Ser Phe	Thr		
	305		310		
Ser His Leu Leu	Gly Asp Ala Gly Ser	Pro Tyr Leu Ile Gly	Phe		
	320		325		
Ile Ser Asp Leu	Ile Arg Gln Ser Thr	Lys Asp Ser Pro Leu	Trp		
	335		340		
Glu Phe Leu Ser	Leu Gly Tyr Ala Leu	Met Leu Cys Pro Phe	Val		
	350		355		
Val Val Leu Gly	Gly Met Phe Phe Leu	Ala Thr Ala Leu Phe	Phe		
	365		370		
Val Ser Asp Arg	Ala Arg Ala Glu Gln	Gln Val Asn Gln Leu	Ala		
	380		385		
Met Pro Pro Ala	Ser Val Lys Val				
	395				

<210> 10
 <211> 553
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 3358383CD1

<400> 10	
Met Ala Phe Gln Asp	Leu Leu Gly His Ala Gly Asp Leu Trp Arg
1	5
Phe Gln Ile Leu Gln	Thr Val Phe Leu Ser Ile Phe Ala Val Ala
	20
Thr Tyr Leu His Phe	Met Leu Glu Asn Phe Thr Ala Phe Ile Pro
	35
Gly His Arg Cys Trp	Val His Ile Leu Asp Asn Asp Thr Val Ser
	50
Asp Asn Asp Thr Gly	Ala Leu Ser Gln Asp Ala Leu Leu Arg Ile
	65
Ser Ile Pro Leu Asp	Ser Asn Met Arg Pro Glu Lys Cys Arg Arg
	80
Phe Val His Pro Gln	Trp Gln Leu Leu His Leu Asn Gly Thr Phe
	95
Pro Asn Thr Ser Asp	Ala Asp Met Glu Pro Cys Val Asp Gly Trp
	100
	105

Val Tyr Asp Arg	110	Ile Ser Phe Ser Ser	115	Thr Ile Val Thr Glu Trp	120
Asp Leu Val Cys	125	Asp Ser Gln Ser Leu	130	Ser Val Ala Lys Phe	135
Val Phe Met Ala	140	Gly Met Met Val Gly	145	Ile Leu Gly Gly His	150
Leu Ser Asp Arg	155	Phe Gly Arg Arg Phe	160	Val Leu Arg Trp Cys Tyr	165
Leu Gln Val Ala	170	Ile Val Gly Thr Cys	175	Ala Leu Ala Pro Thr	180
Phe Leu Ile Tyr	185	Cys Ser Leu Arg Phe	190	Leu Ser Gly Ile Ala Ala	195
Met Ser Leu Ile	200	Thr Asn Thr Ile Met	205	Leu Ile Ala Glu Trp Ala	210
Thr His Arg Phe	215	Gln Ala Met Gly Ile	220	Thr Leu Gly Met Cys Pro	225
Ser Gly Ile Ala	230	Phe Met Thr Leu Ala	235	Gly Leu Ala Phe Ala Ile	240
Arg Asp Trp His	245	Ile Leu Gln Leu Val	250	Val Ser Val Pro Tyr Phe	255
Val Ile Phe Leu	260	Thr Ser Ser Trp Leu	265	Leu Glu Ser Ala Arg Trp	270
Leu Ile Ile Asn	275	Asn Lys Pro Glu Glu	280	Leu Lys Glu Leu Arg	285
Lys Ala Ala His	290	Arg Ser Gly Met Lys	295	Asn Ala Arg Asp Thr Leu	300
Thr Leu Glu Ile	305	Leu Lys Ser Thr Met	310	Lys Glu Leu Glu Ala	315
Ala Gln Lys Lys	320	Lys Pro Ser Leu Cys	325	Glu Met Leu His Met Pro	330
Asn Ile Cys Lys	335	Arg Ile Ser Leu Leu	340	Ser Phe Thr Arg Phe Ala	345
Asn Phe Met Ala	350	Tyr Phe Gly Leu Asn	355	Leu His Val Gln His Leu	360
Gly Asn Asn Val	365	Phe Leu Leu Gln Thr	370	Leu Phe Gly Ala Val Ile	375
Leu Leu Ala Asn	380	Cys Val Ala Pro Trp	385	Ala Leu Lys Tyr Met Thr	390
Arg Arg Ala Ser	395	Gln Met Arg Leu Met	400	Tyr Leu Leu Ala Ile Cys	405
Phe Met Ala Ile	410	Ile Phe Val Pro Gln	415	Glu Met Gln Thr Leu Arg	420
Glu Val Leu Ala	425	Thr Leu Gly Leu Gly	430	Ala Ser Ala Leu Thr Asn	435
Thr Leu Ala Phe	440	Ala His Gly Asn Glu	445	Val Ile Pro Thr Ile Ile	450
Arg Ala Arg Ala	455	Met Gly Ile Asn Ala	460	Thr Phe Ala Asn Ile Ala	465
Gly Ala Leu Ala	470	Pro Leu Met Met Ile	475	Leu Ser Val Tyr Ser Pro	480
Pro Leu Pro Trp	485	Ile Ile Tyr Gly Val	490	Phe Pro Phe Ile Ser Gly	495
Phe Ala Phe Leu	500	Leu Leu Pro Glu Thr	505	Arg Asn Lys Pro Leu Phe	510
Asp Thr Ile Gln	515	Asp Glu Lys Asn Glu	520	Arg Lys Asp Pro Arg Glu	525
Pro Lys Gln Glu	530	Asp Pro Arg Val Glu	535	Val Thr Gln Phe	540
	545		550		

<210> 11
 <211> 213
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature

<223> Incyte ID No: 4250091CD1

<400> 11

Met	Ser	Ser	Gln	Glu	Leu	Val	Thr	Leu	Asn	Val	Gly	Gly	Lys	Ile
1				5					10					15
Phe	Thr	Thr	Arg	Phe	Ser	Thr	Ile	Lys	Gln	Phe	Pro	Ala	Ser	Arg
				20					25					30
Leu	Ala	Arg	Met	Leu	Asp	Gly	Arg	Asp	Gln	Glu	Phe	Lys	Met	Val
				35					40					45
Gly	Gly	Gln	Ile	Phe	Val	Asp	Arg	Asp	Gly	Asp	Leu	Phe	Ser	Phe
				50					55					60
Ile	Leu	Asp	Phe	Leu	Arg	Thr	His	Gln	Leu	Leu	Leu	Pro	Thr	Glu
				65					70					75
Phe	Ser	Asp	Tyr	Leu	Arg	Leu	Gln	Arg	Glu	Ala	Leu	Phe	Tyr	Glu
				80					85					90
Leu	Arg	Ser	Leu	Val	Asp	Leu	Leu	Asn	Pro	Tyr	Leu	Leu	Gln	Pro
				95					100					105
Arg	Pro	Ala	Leu	Val	Glu	Val	His	Phe	Leu	Ser	Arg	Asn	Thr	Gln
				110					115					120
Ala	Phe	Phe	Arg	Val	Phe	Gly	Ser	Cys	Ser	Lys	Thr	Ile	Glu	Met
				125					130					135
Leu	Thr	Gly	Arg	Ile	Thr	Val	Phe	Thr	Glu	Gln	Pro	Ser	Ala	Pro
				140					145					150
Thr	Trp	Asn	Gly	Asn	Phe	Phe	Pro	Pro	Gln	Met	Thr	Leu	Leu	Pro
				155					160					165
Leu	Pro	Pro	Gln	Arg	Pro	Ser	Tyr	His	Asp	Leu	Val	Phe	Gln	Cys
				170					175					180
Gly	Ser	Asp	Ser	Thr	Thr	Asp	Asn	Gln	Thr	Gly	Val	Arg	Tyr	Phe
				185					190					195
Val	Leu	Cys	Ser	Ile	Ser	Leu	Val	Tyr	Gln	Phe	Val	Met	Phe	Ser
				200					205					210
Leu	Lys	Thr												

<210> 12

<211> 476

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 70064803CD1

<400> 12

Met	Ala	Gly	Ser	Asp	Thr	Ala	Pro	Phe	Leu	Ser	Gln	Ala	Asp	Asp
1				5					10					15
Pro	Asp	Asp	Gly	Pro	Val	Pro	Gly	Thr	Pro	Gly	Leu	Pro	Gly	Ser
				20					25					30
Thr	Gly	Asn	Pro	Lys	Ser	Glu	Glu	Pro	Glu	Val	Pro	Asp	Gln	Glu
				35					40					45
Gly	Leu	Gln	Arg	Ile	Thr	Gly	Leu	Ser	Pro	Gly	Arg	Ser	Ala	Leu
				50					55					60
Ile	Val	Ala	Val	Leu	Cys	Tyr	Ile	Asn	Leu	Leu	Asn	Tyr	Met	Asp
				65					70					75
Arg	Phe	Thr	Val	Ala	Gly	Val	Leu	Pro	Asp	Ile	Glu	Gln	Phe	Phe
				80					85					90
Asn	Ile	Gly	Asp	Ser	Ser	Ser	Gly	Leu	Ile	Gln	Thr	Val	Phe	Ile
				95					100					105
Ser	Ser	Tyr	Met	Val	Leu	Ala	Pro	Val	Phe	Gly	Tyr	Leu	Gly	Asp
				110					115					120
Arg	Tyr	Asn	Arg	Lys	Tyr	Leu	Met	Cys	Gly	Gly	Ile	Ala	Phe	Trp
				125					130					135
Ser	Leu	Val	Thr	Leu	Gly	Ser	Ser	Phe	Ile	Pro	Gly	Glu	His	Phe
				140					145					150
Trp	Leu	Leu	Leu	Leu	Thr	Arg	Gly	Leu	Val	Gly	Val	Gly	Glu	Ala
				155					160					165
Ser	Tyr	Ser	Thr	Ile	Ala	Pro	Thr	Leu	Ile	Ala	Asp	Leu	Phe	Val

Ala Asp Gln Arg	170	Arg Met Leu Ser	175	Ile Phe Tyr Phe Ala	180
Pro Val Gly Ser	185	Leu Gly Tyr Ile	190	Ala Gly Ser Lys Val	195
Asp Met Ala Gly	200	Trp His Trp Ala	205	Leu Arg Val Thr Pro	210
Leu Gly Val Val	215	Val Leu Leu Leu	220	Phe Leu Val Val Arg	225
Pro Pro Arg Gly	230	Val Glu Arg His	235	Ser Asp Leu Pro Pro	240
Asn Pro Thr Ser	245	Trp Trp Ala Asp	250	Leu Arg Ala Arg Asn	255
Leu Ile Phe Gly	260	Ile Thr Cys Leu	265	Thr Gly Val Leu Gly	270
Gly Leu Gly Val	275	Glu Ile Ser Arg	280	Leu Arg His Ser Asn	285
Arg Ala Asp Pro	290	Val Cys Ala Thr	295	Gly Leu Leu Gly Ser	300
Pro Phe Leu Phe	305	Leu Ser Leu Ala	310	Cys Ala Arg Gly Ser	315
Ala Thr Tyr Ile	320	Phe Ile Phe Ile	325	Glu Thr Leu Leu Ser	330
Asn Trp Ala Ile	335	Val Ala Asp Ile	340	Leu Tyr Val Val Ile	345
Thr Arg Arg Ser	350	Thr Ala Glu Ala	355	Phe Gln Ile Val Leu	360
Leu Leu Gly Asp	365	Ala Gly Ser Pro	370	Tyr Leu Ile Gly Leu	375
Asp Arg Leu Arg	380	Arg Asn Trp Pro	385	Pro Ser Phe Leu Ser	390
Arg Ala Leu Gln	395	Phe Ser Leu Met	400	Leu Cys Ala Phe Val	405
Leu Gly Gly Ala	410	Ala Phe Leu Gly	415	Thr Ala Ile Phe Ile	420
Asp Arg Arg Arg	425	Ala Gln Leu His	430	Val Gln Gly Leu Leu	435
Ala Gly Ser Thr	440	Asp Asp Arg Ile	445	Val Pro Gln Arg Gly	450
Ser Thr Arg Val	455	Pro Val Ala Ser	460	Val Leu Ile	465
	470		475		

<210> 13

<211> 246

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 70356768CD1

<400> 13

Met Leu His Ala	1	Leu Leu Arg Ser	5	Arg Met Ile Gln	10	Gly Arg Ile	15
Leu Leu Leu Thr	20	Ile Cys Ala Ala	25	Gly Ile Gly Gly	30	Thr Phe Gln	35
Phe Gly Tyr Asn	35	Leu Ser Ile Ile	40	Asn Ala Pro Thr	45	Leu His Ile	50
Gln Glu Phe Thr	50	Asn Glu Thr Trp	55	Gln Ala Arg Thr	60	Gly Glu Pro	65
Leu Pro Asp His	65	Leu Val Leu Leu	70	Met Trp Ser Leu	75	Ile Val Ser	80
Leu Tyr Pro Leu	80	Gly Gly Leu Phe	85	Gly Ala Leu Leu	90	Ala Gly Pro	95
Leu Ala Ile Thr	95	Leu Gly Arg Lys	100	Lys Ser Leu Leu	105	Val Asn Asn	110
Ile Phe Val Val		Ser Ala Ala Ile		Leu Phe Gly Phe		Ser Arg Lys	

Ala Gly Ser Phe	110	Met Ile Met Leu	115	Arg Leu Leu Val	120
Val Asn Ala Gly	125	Val Ser Met Asn Ile	130	Gln Pro Met Tyr Leu	135
Glu Ser Ala Pro	140	Lys Glu Leu Arg Gly	145	Val Ala Met Ser	150
Ala Ile Phe Thr	155	Ala Leu Gly Ile Val	160	Met Gly Gln Val Val	165
Leu Arg Glu Leu	170	Leu Gly Gly Pro Gln	175	Ala Trp Pro Leu Leu	180
Ala Ser Cys Leu	185	Val Pro Gly Ala Leu	190	Gln Leu Ala Ser Leu	195
Leu Leu Pro Glu	200	Ser Pro Arg Tyr Leu	205	Leu Ile Asp Cys Gly	210
Thr Glu Ala Cys	215	Leu Ala Glu Thr Gly	220	Ser Arg Leu Ser Arg	225
Glu Cys Cys Gly	230	Cys Ser	235		240
	245				

<210> 14

<211> 436

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 5674114CD1

<400> 14

Met Gly Leu Ala Arg	Ala Leu Arg Arg	Leu Ser Gly Ala Leu Asp
1	5	10
Ser Gly Asp Ser Arg	Ala Gly Asp Glu	Glu Glu Ala Gly Pro Gly
	20	25
Leu Cys Arg Asn Gly	Trp Ala Pro Ala	Pro Val Gln Ser Pro Val
	35	40
Gly Arg Arg Arg Gly	Arg Phe Val Lys	Lys Asp Gly His Cys Asn
	50	55
Val Arg Phe Val Asn	Leu Gly Gly Gln	Gly Ala Arg Tyr Leu Ser
	65	70
Asp Leu Phe Thr Thr	Cys Val Asp Val	Arg Trp Arg Trp Met Cys
	80	85
Leu Leu Phe Ser Cys	Ser Phe Leu Ala	Ser Trp Leu Leu Phe Gly
	95	100
Leu Ala Phe Trp Leu	Ile Ala Ser Leu	His Gly Asp Leu Ala Ala
	110	115
Pro Pro Pro Pro Ala	Pro Cys Phe Ser	His Val Ala Ser Phe Leu
	125	130
Ala Ala Phe Leu Phe	Ala Leu Glu Thr	Gln Thr Ser Ile Gly Tyr
	140	145
Gly Val Arg Ser Val	Thr Glu Glu Cys	Pro Ala Ala Val Ala Ala
	155	160
Val Val Leu Gln Cys	Ile Ala Gly Cys	Val Leu Asp Ala Phe Val
	170	175
Val Gly Ala Val Met	Ala Lys Met Ala	Lys Pro Lys Lys Arg Asn
	185	190
Glu Thr Leu Val Phe	Ser Glu Asn Ala	Val Val Ala Leu Arg Asp
	200	205
His Arg Leu Cys Leu	Met Trp Arg Val	Gly Asn Leu Arg Arg Ser
	215	220
His Leu Val Glu Ala	His Val Arg Ala	Gln Leu Leu Gln Pro Arg
	230	235
Val Thr Pro Glu Gly	Glu Tyr Ile Pro	Leu Asp His Gln Asp Val
	245	250
Asp Val Gly Phe Asp	Gly Gly Thr Asp	Arg Ile Phe Leu Val Ser
	260	265
Pro Ile Thr Ile Val	His Glu Ile Asp	Ser Ala Ser Pro Leu Tyr

	275		280		285
Glu Leu Gly Arg	Ala Glu Leu Ala Arg	Ala Asp Phe Glu Leu Val			
	290		295		300
Val Ile Leu Glu	Gly Met Val Glu Ala Thr	Ala Met Thr Thr Gln			
	305		310		315
Cys Arg Ser Ser	Tyr Leu Pro Gly Glu	Leu Leu Trp Gly His Arg			
	320		325		330
Phe Glu Pro Val	Leu Phe Gln Arg Gly	Ser Gln Tyr Glu Val Asp			
	335		340		345
Tyr Arg His Phe	His Arg Thr Tyr Glu	Val Pro Gly Thr Pro Val			
	350		355		360
Cys Ser Ala Lys	Glu Leu Asp Glu Arg	Ala Glu Gln Ala Ser His			
	365		370		375
Ser Leu Lys Ser	Ser Phe Pro Gly Ser	Leu Thr Ala Phe Cys Tyr			
	380		385		390
Glu Asn Glu Leu	Ala Leu Ser Cys Cys	Gln Glu Glu Asp Glu Asp			
	395		400		405
Asp Glu Thr Glu	Glu Gly Asn Gly Val	Glu Thr Glu Asp Gly Ala			
	410		415		420
Ala Ser Pro Arg	Val Leu Thr Pro Thr	Leu Ala Leu Thr Leu Pro			
	425		430		435
Pro					

<210> 15
 <211> 453
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 1254635CD1

<400> 15
 Met Leu Lys Met Val Leu Thr Glu Asn Pro Asn Gln Glu Ile Ala
 1 5 10 15
 Thr Ser Leu Glu Phe Leu Leu Leu Gln Asn Ser Pro Gly Ser Leu
 20 25 30
 Arg Ala Gln Gln Arg Met Ser Tyr Tyr Gly Ser Ser Tyr His Ile
 35 40 45
 Ile Asn Ala Asp Ala Lys Tyr Pro Gly Tyr Pro Pro Glu His Ile
 50 55 60
 Ile Ala Glu Lys Arg Arg Ala Arg Arg Leu Leu His Lys Asp
 65 70 75
 Gly Ser Cys Asn Val Tyr Phe Lys His Ile Phe Gly Glu Trp Gly
 80 85 90
 Ser Tyr Val Val Asp Ile Phe Thr Thr Leu Val Asp Thr Lys Trp
 95 100 105
 Arg His Met Phe Val Ile Phe Ser Leu Ser Tyr Ile Leu Ser Trp
 110 115 120
 Leu Ile Phe Gly Ser Val Phe Trp Leu Ile Ala Phe His His Gly
 125 130 135
 Asp Leu Leu Asn Asp Pro Asp Ile Thr Pro Cys Val Asp Asn Val
 140 145 150
 His Ser Phe Thr Gly Ala Phe Leu Phe Ser Leu Glu Thr Gln Thr
 155 160 165
 Thr Ile Gly Tyr Gly Tyr Arg Cys Val Thr Glu Glu Cys Ser Val
 170 175 180
 Ala Val Leu Met Val Ile Leu Gln Ser Ile Leu Ser Cys Ile Ile
 185 190 195
 Asn Thr Phe Ile Ile Gly Ala Ala Leu Ala Lys Met Ala Thr Ala
 200 205 210
 Arg Lys Arg Ala Gln Thr Ile Arg Phe Ser Tyr Phe Ala Leu Ile
 215 220 225
 Gly Met Arg Asp Gly Lys Leu Cys Leu Met Trp Arg Ile Gly Asp
 230 235 240
 Phe Arg Pro Asn His Val Val Glu Gly Thr Val Arg Ala Gln Leu

Leu	Arg	Tyr	Thr	245	Asp	Ser	Glu	Gly	250	Arg	Met	Thr	Met	Ala	Phe	255
				260					265							270
Lys	Asp	Leu	Lys	275	Leu	Val	Asn	Asp	280	Ile	Ile	Leu	Val	Thr	Pro	285
Val	Thr	Ile	Val	290	His	Glu	Ile	Asp	295	His	Ser	Pro	Leu	Tyr	Ala	300
Leu	Asp	Arg	Lys	305	Ala	Val	Ala	Lys	310	Asn	Phe	Glu	Ile	Leu	Val	315
Thr	Phe	Ile	Tyr	320	Thr	Gly	Asp	Ser	325	Gly	Thr	Ser	His	Gln	Ser	330
Arg	Ser	Ser	Tyr	335	Val	Pro	Arg	Glu	340	Leu	Trp	Gly	His	Arg	Phe	345
Asn	Asp	Val	Leu	350	Glu	Val	Lys	Arg	355	Tyr	Tyr	Lys	Val	Asn	Cys	360
Leu	Gln	Phe	Glu	365	Gly	Ser	Val	Glu	370	Tyr	Ala	Pro	Phe	Cys	Ser	375
Ala	Lys	Gln	Leu	380	Asp	Trp	Lys	Asp	385	Gln	Leu	His	Ile	Glu	Lys	390
Ala	Pro	Pro	Val	395	Arg	Glu	Ser	Cys	400	Ser	Asp	Thr	Lys	Ala	Arg	405
Arg	Arg	Ser	Phe	410	Ser	Ala	Val	Ala	415	Val	Ser	Ser	Cys	Glu	Asn	420
Pro	Glu	Glu	Thr	425	Thr	Thr	Ser	Ala	430	His	Glu	Tyr	Arg	Glu	Thr	435
Pro	Tyr	Gln	Lys	440	Ala	Leu	Leu	Thr	445	Asn	Arg	Ile	Ser	Val	Glu	450
Ser	Gln	Met														

<210> 16

<211> 299

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1670595CD1

<400> 16

Met	Ala	Ser	Glu	Ser	Ser	Pro	Leu	Leu	Ala	Tyr	Arg	Leu	Leu	Gly	
1				5					10					15	
Glu	Glu	Gly	Val	Ala	Leu	Pro	Ala	Asn	Gly	Ala	Gly	Gly	Pro	Gly	
				20					25					30	
Gly	Ala	Ser	Ala	Arg	Lys	Leu	Ser	Thr	Phe	Leu	Gly	Val	Val	Val	
				35					40					45	
Pro	Thr	Val	Leu	Ser	Met	Phe	Ser	Ile	Val	Val	Phe	Leu	Arg	Ile	
				50					55					60	
Gly	Phe	Val	Val	Gly	His	Ala	Gly	Leu	Leu	Gln	Ala	Leu	Ala	Met	
				65					70					75	
Leu	Leu	Val	Ala	Tyr	Phe	Ile	Leu	Ala	Leu	Thr	Val	Leu	Ser	Val	
				80					85					90	
Cys	Ala	Ile	Ala	Thr	Asn	Gly	Ala	Val	Gln	Gly	Gly	Gly	Ala	Tyr	
				95					100					105	
Cys	Ile	Leu	Gln	His	Arg	Trp	Thr	Gly	Met	Pro	Gln	Gly	Pro	Val	
				110					115					120	
Gly	Ser	Gly	Ser	Cys	Pro	Arg	Ala	Thr	Ala	Trp	Asn	Leu	Leu	Tyr	
				125					130					135	
Gly	Ser	Leu	Leu	Leu	Gly	Leu	Val	Gly	Gly	Val	Cys	Thr	Leu	Gly	
				140					145					150	
Ala	Gly	Leu	Tyr	Ala	Arg	Ala	Ser	Phe	Leu	Thr	Phe	Leu	Leu	Val	
				155					160					165	
Ser	Gly	Ser	Leu	Ala	Ser	Val	Leu	Ile	Ser	Phe	Val	Ala	Val	Gly	
				170					175					180	
Pro	Arg	Asp	Ile	Arg	Leu	Thr	Pro	Arg	Pro	Gly	Pro	Asn	Gly	Ser	
				185					190					195	
Ser	Leu	Pro	Pro	Arg	Phe	Gly	His	Phe	Thr	Gly	Phe	Asn	Ser	Ser	

Thr	Leu	Lys	Asp	Asn	Leu	Gly	Ala	Gly	Tyr	Ala	Glu	Asp	Tyr	Thr	200	205	210
Thr	Gly	Ala	Val	Met	Asn	Phe	Ala	Ser	Val	Phe	Ala	Val	Leu	Phe	215	220	225
Asn	Gly	Arg	His	His	Gly	Trp	Gly	Gln	His	Val	Arg	Gly	Ala	Glu	230	235	240
Gly	Pro	Gln	Pro	Gly	Asp	Pro	Ser	Gly	His	Asp	Arg	Arg	Arg	Arg	245	250	255
Leu	His	Leu	Leu	Arg	Leu	Cys	Pro	Ala	Phe	Leu	Ser	Leu	Gln	Pro	260	265	270
Pro	Phe	Thr	Gly	Ala	Leu	Met	Leu	Gly	Ala	Arg	Pro	Pro	Leu		275	280	285
															290	295	

<210> 17
 <211> 606
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 1859560CD1

<400> 17

Met	Pro	Ser	Ser	Val	Thr	Ala	Leu	Gly	Gln	Ala	Arg	Ser	Ser	Gly	1	5	10	15
Pro	Gly	Met	Ala	Pro	Ser	Ala	Cys	Cys	Cys	Ser	Pro	Ala	Ala	Leu	20	25	30	35
Gln	Arg	Arg	Leu	Pro	Ile	Leu	Ala	Trp	Leu	Pro	Ser	Tyr	Ser	Leu	40	45	50	55
Gln	Trp	Leu	Lys	Met	Asp	Phe	Val	Ala	Gly	Leu	Ser	Val	Gly	Leu	60	65	70	75
Thr	Ala	Ile	Pro	Gln	Ala	Leu	Ala	Tyr	Ala	Glu	Val	Ala	Gly	Leu	80	85	90	95
Pro	Pro	Gln	Tyr	Gly	Leu	Tyr	Ser	Ala	Phe	Met	Gly	Cys	Phe	Val	100	105	110	115
Tyr	Phe	Phe	Leu	Gly	Thr	Ser	Arg	Asp	Val	Thr	Leu	Gly	Pro	Thr	120	125	130	135
Ala	Ile	Met	Ser	Leu	Leu	Val	Ser	Phe	Tyr	Thr	Phe	His	Glu	Pro	140	145	150	155
Ala	Tyr	Ala	Val	Leu	Leu	Ala	Phe	Leu	Ser	Gly	Cys	Ile	Gln	Leu	160	165	170	175
Ala	Met	Gly	Val	Leu	Arg	Leu	Gly	Phe	Leu	Leu	Asp	Phe	Ile	Ser	180	185	190	195
Tyr	Pro	Val	Ile	Lys	Gly	Phe	Thr	Ser	Ala	Ala	Ala	Val	Thr	Ile	200	205	210	215
Gly	Phe	Gly	Gln	Ile	Lys	Asn	Leu	Leu	Gly	Leu	Gln	Asn	Ile	Pro	220	225	230	235
Arg	Pro	Phe	Phe	Leu	Gln	Val	Tyr	His	Thr	Phe	Leu	Arg	Ile	Ala	240	245	250	255
Glu	Thr	Arg	Val	Gly	Asp	Ala	Val	Leu	Gly	Leu	Val	Cys	Met	Leu	260	265	270	275
Leu	Leu	Leu	Val	Leu	Lys	Leu	Met	Arg	Asp	His	Val	Pro	Pro	Val	280	285	290	295
His	Pro	Glu	Met	Pro	Pro	Gly	Val	Arg	Leu	Ser	Arg	Gly	Leu	Val	300	305	310	315
Trp	Ala	Ala	Thr	Thr	Ala	Arg	Asn	Ala	Leu	Val	Val	Ser	Phe	Ala				
Ala	Leu	Val	Ala	Tyr	Ser	Phe	Glu	Val	Thr	Gly	Tyr	Gln	Pro	Phe				
Ile	Leu	Thr	Gly	Glu	Thr	Ala	Glu	Gly	Leu	Pro	Pro	Val	Arg	Ile				
Pro	Pro	Phe	Ser	Val	Thr	Thr	Ala	Asn	Gly	Thr	Ile	Ser	Phe	Thr				
Glu	Met	Val	Gln	Asp	Met	Gly	Ala	Gly	Leu	Ala	Val	Val	Pro	Leu				
Met	Gly	Leu	Leu	Glu	Ser	Ile	Ala	Val	Ala	Lys	Ala	Phe	Ala	Ser				

Gln Asn Asn Tyr	320	Arg Ile Asp Ala Asn	325	Gln Glu Leu Leu Ala	330
	335		340		345
Gly Leu Thr Asn Met	350	Leu Gly Ser Leu Val	355	Ser Ser Tyr Pro	360
	365		370		375
Thr Gly Ser Phe Gly	380	Arg Thr Ala Val Asn	385	Ala Gln Ser Gly	390
	395		400		405
Cys Thr Pro Ala Gly	410	Gly Gly Leu Val Thr	415	Val Leu Val Leu	420
	425		430		435
Ser Leu Asp Tyr Leu	440	Thr Ser Leu Phe Tyr	445	Tyr Tyr Ile Pro Lys	450
	455		460		465
Ala Leu Ala Ala Val	470	Ile Ile Met Ala Val	475	Ala Pro Leu Phe	480
	485		490		495
Thr Lys Ile Phe Arg	500	Thr Leu Trp Arg Val	505	Lys Arg Leu Asp	510
	515		520		525
Leu Pro Leu Cys Val	530	Thr Phe Leu Leu Cys	535	Phe Trp Glu Val	540
	545		550		555
Tyr Gly Ile Leu Ala	560	Gly Ala Leu Val Ser	565	Leu Leu Met Leu	570
	575		580		585
His Ser Ala Ala Arg	590	Pro Glu Thr Lys Val	595	Ser Glu Gly Pro	600
	605				
Leu Val Leu Gln Pro		Ala Ser Gly Leu Ser		Phe Pro Ala Met	
Ala Leu Arg Glu Glu		Ile Leu Ser Arg Ala		Leu Glu Val Ser	
Pro Arg Cys Leu Val		Leu Glu Cys Thr His		Val Cys Ser Ile	
Tyr Thr Val Val Leu		Gly Leu Gly Glu Leu		Leu Gln Asp Phe	
Lys Gln Gly Val Ala		Leu Ala Phe Val Gly		Leu Gln Val Pro	
Leu Arg Val Leu Leu		Ser Ala Asp Leu Lys		Gly Phe Gln Tyr	
Ser Thr Leu Glu Glu		Ala Glu Lys His Leu		Arg Gln Glu Pro	
Thr Gln Pro Tyr Asn		Ile Arg Glu Asp Ser		Ile Leu Asp Gln	
Val Ala Leu Leu Lys		Ala			

<210> 18
 <211> 324
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No: 5530164CD1

<400> 18

Met Ser Val Glu Asp	Gly Gly Met Pro Gly	Leu Gly Arg Pro Arg
1	5	10
Gln Ala Arg Trp Thr	Leu Met Leu Leu Leu	Ser Thr Ala Met Tyr
	20	25
Gly Ala His Ala Pro	Leu Leu Ala Leu Cys	His Val Asp Gly Arg
	35	40
Val Pro Phe Arg Pro	Ser Ser Ala Val Leu	Leu Thr Glu Leu Thr
	50	55
Lys Leu Leu Leu Cys	Ala Phe Ser Leu Leu	Val Gly Trp Gln Ala
	65	70
Trp Pro Gln Gly Pro	Pro Pro Trp Arg Gln	Ala Ala Pro Phe Ala
	80	85
Leu Ser Ala Leu Leu	Tyr Gly Ala Asn Asn	Asn Leu Val Ile Tyr
	95	100
Leu Gln Arg Tyr Met	Asp Pro Ser Thr Tyr	Gln Val Leu Ser Asn
	110	115
Leu Lys Ile Gly Ser	Thr Ala Val Leu Tyr	Cys Leu Cys Leu Arg

	125		130		135
His Arg Leu Ser	Val Arg Gln Gly Leu	Ala Leu Leu Leu Leu	Met		
	140		145		150
Ala Ala Gly Ala	Cys Tyr Ala Ala Gly	Gly Leu Gln Val Pro	Gly		
	155		160		165
Asn Thr Leu Pro	Ser Pro Pro Pro Ala	Ala Ala Ala Ser Pro	Met		
	170		175		180
Pro Leu His Ile	Thr Pro Leu Gly Leu	Leu Leu Leu Ile Leu	Tyr		
	185		190		195
Cys Leu Ile Ser	Gly Leu Ser Ser Val	Tyr Thr Glu Leu Leu	Met		
	200		205		210
Lys Arg Gln Arg	Leu Pro Leu Ala Leu	Gln Asn Leu Phe Leu	Tyr		
	215		220		225
Thr Phe Gly Val	Leu Leu Asn Leu Gly	Leu His Ala Gly Gly	Gly		
	230		235		240
Ser Gly Pro Gly	Leu Leu Glu Gly Phe	Ser Gly Trp Ala Ala	Leu		
	245		250		255
Val Val Leu Ser	Gln Ala Leu Asn Gly	Leu Leu Met Ser Ala	Val		
	260		265		270
Met Lys His Gly	Ser Ser Ile Thr Arg	Leu Phe Val Val Ser	Cys		
	275		280		285
Ser Leu Val Val	Asn Ala Val Leu Ser	Ala Val Leu Leu Arg	Leu		
	290		295		300
Gln Leu Thr Ala	Ala Phe Phe Leu Ala	Thr Leu Leu Ile Gly	Leu		
	305		310		315
Ala Met Arg Leu	Tyr Tyr Gly Ser Arg				
	320				

<210> 19

<211> 445

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 139115CD1

<400> 19

Met Thr Leu Thr Gly	Pro Leu Thr Thr Gln	Tyr Val Tyr Arg Arg	
1	5	10	15
Ile Trp Glu Glu Thr	Gly Asn Tyr Thr Phe	Ser Ser Asp Ser Asn	
	20	25	30
Ile Ser Glu Cys Glu	Lys Asn Lys Ser Ser	Pro Ile Phe Ala Phe	
	35	40	45
Gln Glu Glu Val Gln	Lys Lys Val Ser Arg	Phe Asn Leu Gln Met	
	50	55	60
Asp Ile Ser Gly Leu	Ile Pro Gly Leu Val	Ser Thr Phe Ile Leu	
	65	70	75
Leu Ser Ile Ser Asp	His Tyr Gly Arg Lys	Phe Pro Met Ile Leu	
	80	85	90
Ser Ser Val Gly Ala	Leu Ala Thr Ser Val	Trp Leu Cys Leu Leu	
	95	100	105
Cys Tyr Phe Ala Phe	Pro Phe Gln Leu Leu	Ile Ala Ser Thr Phe	
	110	115	120
Ile Gly Ala Phe Cys	Gly Asn Tyr Thr Thr	Phe Trp Gly Ala Cys	
	125	130	135
Phe Ala Tyr Ile Val	Asp Gln Cys Lys Glu	His Lys Gln Lys Thr	
	140	145	150
Ile Arg Ile Ala Ile	Ile Asp Phe Leu Leu	Gly Leu Val Thr Gly	
	155	160	165
Leu Thr Gly Leu Ser	Ser Gly Tyr Phe Ile	Arg Glu Leu Gly Phe	
	170	175	180
Glu Trp Ser Phe Leu	Ile Ile Ala Val Ser	Leu Ala Val Asn Leu	
	185	190	195
Ile Tyr Ile Leu Phe	Phe Leu Gly Asp Pro	Val Lys Glu Cys Ser	
	200	205	210
Ser Gln Asn Val Thr	Met Ser Cys Ser Glu	Gly Phe Lys Asn Leu	

Phe	Tyr	Arg	Thr	215	Met	Leu	Phe	Lys	220	Asn	Ala	Ser	Gly	Lys	225
				230					235						240
Arg	Phe	Leu	Leu	245	Cys	Leu	Leu	Leu	250	Thr	Val	Ile	Thr	Tyr	255
				260					265						270
Phe	Val	Val	Ile	275	Gly	Ile	Ala	Pro	280	Phe	Ile	Leu	Tyr	Glu	285
				290					295						300
Asp	Ser	Pro	Leu	305	Cys	Trp	Asn	Glu	310	Phe	Ile	Gly	Tyr	Gly	315
				320					325						330
Ala	Leu	Gly	Ser	335	Ala	Ser	Phe	Leu	340	Ser	Phe	Leu	Gly	Ile	345
				350					355						360
Leu	Phe	Ser	Tyr	365	Cys	Met	Glu	Asp	370	Ile	His	Met	Ala	Phe	375
				380					385						390
Ile	Phe	Thr	Thr	395	Met	Thr	Gly	Met	400	Met	Thr	Ala	Phe	Ala	405
				410					415						420
Thr	Thr	Leu	Met	425	Met	Phe	Leu	Ala	430	Val	Pro	Phe	Leu	Phe	435
				440					445						
Ile	Val	Pro	Phe		Ser	Val	Leu	Arg		Met	Leu	Ser	Lys	Val	
Arg	Ser	Thr	Glu		Gln	Gly	Thr	Leu		Ala	Cys	Ile	Ala	Phe	
Glu	Thr	Leu	Gly		Gly	Val	Thr	Ala		Ser	Thr	Phe	Asn	Gly	
Tyr	Ser	Ala	Thr		Val	Ala	Trp	Tyr		Gly	Phe	Thr	Phe	Leu	
Ser	Ala	Gly	Leu		Leu	Leu	Pro	Ala		Ile	Ser	Leu	Cys	Val	
Lys	Cys	Thr	Ser		Trp	Asn	Glu	Gly		Ser	Tyr	Glu	Leu	Leu	
Glu	Glu	Ser	Ser		Glu	Asp	Ala	Ser		Asp	Arg				

<210> 20

<211> 337

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1702940CD1

<400> 20

Met	Asn	Pro	Glu	Ser	Ser	Ile	Phe	Ile	Glu	Asp	Tyr	Leu	Lys	Tyr
1				5					10					15
Phe	Gln	Asp	Gln	Val	Ser	Arg	Glu	Asn	Leu	Leu	Gln	Leu	Leu	Thr
				20					25					30
Asp	Asp	Glu	Ala	Trp	Asn	Gly	Phe	Val	Ala	Ala	Ala	Glu	Leu	Pro
				35					40					45
Arg	Asp	Glu	Ala	Asp	Glu	Leu	Arg	Lys	Ala	Leu	Asn	Lys	Leu	Ala
				50					55					60
Ser	His	Met	Val	Met	Lys	Asp	Lys	Asn	Arg	His	Asp	Lys	Asp	Gln
				65					70					75
Gln	His	Arg	Gln	Trp	Phe	Leu	Lys	Glu	Phe	Pro	Arg	Leu	Lys	Arg
				80					85					90
Glu	Leu	Glu	Asp	His	Ile	Arg	Lys	Leu	Arg	Ala	Leu	Ala	Glu	Glu
				95					100					105
Val	Glu	Gln	Val	His	Arg	Gly	Thr	Thr	Ile	Ala	Asn	Val	Val	Ser
				110					115					120
Asn	Ser	Val	Gly	Thr	Thr	Ser	Gly	Ile	Leu	Thr	Leu	Leu	Gly	Leu
				125					130					135
Gly	Leu	Ala	Pro	Phe	Thr	Glu	Gly	Ile	Ser	Phe	Val	Leu	Leu	Asp
				140					145					150
Thr	Gly	Met	Gly	Leu	Gly	Ala	Ala	Ala	Ala	Val	Ala	Gly	Ile	Thr
				155					160					165
Cys	Ser	Val	Val	Glu	Leu	Val	Asn	Lys	Leu	Arg	Ala	Arg	Ala	Gln
				170					175					180
Ala	Arg	Asn	Leu	Asp	Gln	Ser	Gly	Thr	Asn	Val	Ala	Lys	Val	Met

Lys	Glu	Phe	Val	Gly	Gly	Asn	Thr	Pro	Asn	Val	Leu	Thr	Leu	Val	185	190	195
Asp	Asn	Trp	Tyr	Gln	Val	Thr	Gln	Gly	Ile	Gly	Arg	Asn	Ile	Arg	200	205	210
Ala	Ile	Arg	Arg	Ala	Arg	Ala	Asn	Pro	Gln	Leu	Gly	Ala	Tyr	Ala	215	220	225
Pro	Pro	Pro	His	Val	Ile	Gly	Arg	Ile	Ser	Ala	Glu	Gly	Gly	Glu	230	235	240
Gln	Val	Glu	Arg	Val	Val	Glu	Gly	Pro	Ala	Gln	Ala	Met	Ser	Arg	245	250	255
Gly	Thr	Met	Ile	Val	Gly	Ala	Ala	Thr	Gly	Gly	Ile	Leu	Leu	Leu	260	265	270
Leu	Asp	Val	Val	Ser	Leu	Ala	Tyr	Glu	Ser	Lys	His	Leu	Leu	Glu	275	280	285
Gly	Ala	Lys	Ser	Glu	Ser	Ala	Glu	Glu	Leu	Lys	Lys	Arg	Ala	Gln	290	295	300
Glu	Leu	Glu	Gly	Lys	Leu	Asn	Phe	Leu	Thr	Lys	Ile	His	Glu	Met	305	310	315
Leu	Gln	Pro	Gly	Gln	Asp	Gln			325					330	320	325	330
															335		

<210> 21

<211> 273

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1703342CD1

<400> 21

Met	Ala	Thr	Trp	Asp	Glu	Lys	Ala	Val	Thr	Arg	Arg	Ala	Lys	Val	1	5	10	15
Ala	Pro	Ala	Glu	Arg	Met	Ser	Lys	Phe	Leu	Arg	His	Phe	Thr	Val	20	25	30	35
Val	Gly	Asp	Asp	Tyr	His	Ala	Trp	Asn	Ile	Asn	Tyr	Lys	Lys	Trp	40	45	50	55
Glu	Asn	Glu	Glu	Glu	Glu	Glu	Glu	Glu	Glu	Gln	Pro	Pro	Pro	Thr	60	65	70	75
Pro	Val	Ser	Gly	Glu	Glu	Gly	Arg	Ala	Ala	Ala	Pro	Asp	Val	Ala	80	85	90	95
Met	Leu	Arg	Lys	Leu	Phe	Ser	Ser	His	Arg	Phe	Gln	Val	Ile	Ile	100	105	110	115
Ile	Cys	Leu	Val	Val	Leu	Asp	Ala	Leu	Leu	Val	Leu	Ala	Glu	Leu	120	125	130	135
Ile	Leu	Asp	Leu	Lys	Ile	Ile	Gln	Pro	Asp	Lys	Asn	Asn	Tyr	Ala	140	145	150	155
Ala	Met	Val	Phe	His	Tyr	Met	Ser	Ile	Thr	Ile	Leu	Val	Phe	Phe	160	165	170	175
Met	Met	Glu	Ile	Ile	Phe	Lys	Leu	Phe	Val	Phe	Arg	Leu	Glu	Phe	180	185	190	195
Phe	His	His	Lys	Phe	Glu	Ile	Leu	Asp	Ala	Val	Val	Val	Val	Val	200	205	210	215
Ser	Phe	Ile	Leu	Asp	Ile	Val	Leu	Leu	Phe	Gln	Glu	His	Gln	Phe	220	225	230	235
Glu	Ala	Leu	Gly	Leu	Leu	Ile	Leu	Leu	Arg	Leu	Trp	Arg	Val	Ala	240	245	250	255
Arg	Ile	Ile	Asn	Gly	Ile	Ile	Ile	Ser	Val	Lys	Thr	Arg	Ser	Glu	260	265	270	275
Arg	Gln	Leu	Leu	Arg	Leu	Lys	Gln	Met	Asn	Val	Gln	Leu	Ala	Ala	280	285	290	295
Lys	Ile	Gln	His	Leu	Glu	Phe	Ser	Cys	Ser	Glu	Lys	Glu	Gln	Glu	300	305	310	315
Ile	Glu	Arg	Leu	Asn	Lys	Leu	Leu	Arg	Gln	His	Gly	Leu	Leu	Gly	320	325	330	335

Glu Val Asn 260 265 270
 <210> 22
 <211> 710
 <212> PRT
 <213> Homo sapiens
 <220>
 <221> misc_feature
 <223> Incyte ID No: 1727529CD1
 <400> 22
 Met Gly Gly Lys Gln Arg Asp Glu Asp Asp Glu Ala Tyr Gly Lys
 1 5 10 15
 Pro Val Lys Tyr Asp Pro Ser Phe Arg Gly Pro Ile Lys Asn Arg
 20 25 30
 Ser Cys Thr Asp Val Ile Cys Cys Val Leu Phe Leu Leu Phe Ile
 35 40 45
 Leu Gly Tyr Ile Val Val Gly Ile Val Ala Trp Leu Tyr Gly Asp
 50 55 60
 Pro Arg Gln Val Leu Tyr Pro Arg Asn Ser Thr Gly Ala Tyr Cys
 65 70 75
 Gly Met Gly Glu Asn Lys Asp Lys Pro Tyr Leu Leu Tyr Phe Asn
 80 85 90
 Ile Phe Ser Cys Ile Leu Ser Ser Asn Ile Ile Ser Val Ala Glu
 95 100 105
 Asn Gly Leu Gln Cys Pro Thr Pro Gln Val Cys Val Ser Ser Cys
 110 115 120
 Pro Glu Asp Pro Trp Thr Val Gly Lys Asn Glu Phe Ser Gln Thr
 125 130 135
 Val Gly Glu Val Phe Tyr Thr Lys Asn Arg Asn Phe Cys Leu Pro
 140 145 150
 Gly Val Pro Trp Asn Met Thr Val Ile Thr Ser Leu Gln Gln Glu
 155 160 165
 Leu Cys Pro Ser Phe Leu Leu Pro Ser Ala Pro Ala Leu Gly Arg
 170 175 180
 Cys Phe Pro Trp Thr Asn Ile Thr Pro Pro Ala Leu Pro Gly Ile
 185 190 195
 Thr Asn Asp Thr Thr Ile Gln Gln Gly Ile Ser Gly Leu Ile Asp
 200 205 210
 Ser Leu Asn Ala Arg Asp Ile Ser Val Lys Ile Phe Glu Asp Phe
 215 220 225
 Ala Gln Ser Trp Tyr Trp Ile Leu Val Ala Leu Gly Val Ala Leu
 230 235 240
 Val Leu Ser Leu Leu Phe Ile Leu Leu Leu Arg Leu Val Ala Gly
 245 250 255
 Pro Leu Val Leu Val Leu Ile Leu Gly Val Leu Gly Val Leu Ala
 260 265 270
 Tyr Gly Ile Tyr Tyr Cys Trp Glu Glu Tyr Arg Val Leu Arg Asp
 275 280 285
 Lys Gly Ala Ser Ile Ser Gln Leu Gly Phe Thr Thr Asn Leu Ser
 290 295 300
 Ala Tyr Gln Ser Val Gln Glu Thr Trp Leu Ala Ala Leu Ile Val
 305 310 315
 Leu Ala Val Leu Glu Ala Ile Leu Leu Leu Val Leu Ile Phe Leu
 320 325 330
 Arg Gln Arg Ile Arg Ile Ala Ile Ala Leu Leu Lys Glu Ala Ser
 335 340 345
 Lys Ala Val Gly Gln Met Met Ser Thr Met Phe Tyr Pro Leu Val
 350 355 360
 Thr Phe Val Leu Leu Ile Cys Ile Ala Tyr Trp Ala Met Thr
 365 370 375
 Ala Leu Tyr Leu Ala Thr Ser Gly Gln Pro Gln Tyr Val Leu Trp
 380 385 390
 Ala Ser Asn Ile Ser Ser Pro Gly Cys Glu Lys Val Pro Ile Asn

Thr Ser Cys Asn	Pro	Thr Ala His Leu	Val Asn Ser Ser Cys	Pro
Gly Leu Met Cys	Val	Phe Gln Gly Tyr	Ser Ser Lys Gly Leu	Ile
Gln Arg Ser Val	Phe	Asn Leu Gln Ile	Tyr Gly Val Leu Gly	Leu
Phe Trp Thr Leu	Asn	Trp Val Leu Ala	Leu Gly Gln Cys Val	Leu
Ala Gly Ala Phe	Ala	Ser Phe Tyr Trp	Ala Phe His Lys Pro	Gln
Asp Ile Pro Thr	Phe	Pro Leu Ile Ser	Ala Phe Ile Arg Thr	Leu
Arg Tyr His Thr	Gly	Ser Leu Ala Phe	Gly Ala Leu Ile Leu	Thr
Leu Val Gln Ile	Ala	Arg Val Ile Leu	Glu Tyr Ile Asp His	Lys
Leu Arg Gly Val	Gln	Asn Pro Val Ala	Arg Cys Ile Met Cys	Cys
Phe Lys Cys Cys	Leu	Trp Cys Leu Glu	Lys Phe Ile Lys Phe	Leu
Asn Arg Asn Ala	Tyr	Ile Met Ile Ala	Ile Tyr Gly Lys Asn	Phe
Cys Val Ser Ala	Lys	Asn Ala Phe Met	Leu Leu Met Arg Asn	Ile
Val Arg Val Val	Val	Leu Asp Lys Val	Thr Asp Leu Leu Leu	Phe
Phe Gly Lys Leu	Leu	Val Val Gly Gly	Val Gly Val Leu Ser	Phe
Phe Phe Phe Ser	Gly	Arg Ile Pro Gly	Leu Gly Lys Asp Phe	Lys
Ser Pro His Leu	Asn	Tyr Tyr Trp Leu	Pro Ile Met Thr Ser	Ile
Leu Gly Ala Tyr	Val	Ile Ala Ser Gly	Phe Phe Ser Val Phe	Gly
Met Cys Val Asp	Thr	Leu Phe Leu Cys	Phe Leu Glu Asp Leu	Glu
Arg Asn Asn Gly	Ser	Leu Asp Arg Pro	Tyr Tyr Met Ser Lys	Ser
Leu Leu Lys Ile	Leu	Gly Lys Lys Asn	Glu Ala Pro Pro Asp	Asn
Lys Lys Arg Lys	Lys			

<210> 23

<211> 476

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 2289333CD1

<400> 23

Glu Gln Asn Phe Asp	Gly Thr Ser Asp	Glu Glu His Glu Gln Glu
1 5	10	15
Leu Leu Pro Val Gln	Lys His Tyr Gln	Leu Asp Asp Gln Glu Gly
20 25	30	35
Ile Ser Phe Val Gln	Thr Leu Met His	Leu Leu Lys Gly Asn Ile
35 40	45	50
Gly Thr Gly Leu Leu	Gly Leu Pro Leu	Ala Ile Lys Asn Ala Gly
50 55	60	65
Ile Val Leu Gly Pro	Ile Ser Leu Val	Phe Ile Gly Ile Ile Ser
65 70	75	80
Val His Cys Met His	Ile Leu Val Arg	Cys Ser His Phe Leu Cys
80 85	90	95
Leu Arg Phe Lys Lys	Ser Thr Leu Gly Tyr	Ser Asp Thr Val Ser

Phe	Ala	Met	Glu	Val	Ser	Pro	Trp	Ser	Cys	Leu	Gln	Lys	Gln	Ala	95	100	105
Ala	Trp	Gly	Arg	Ser	Val	Val	Asp	Phe	Phe	Leu	Val	Ile	Thr	Gln	110	115	120
Leu	Gly	Phe	Cys	Ser	Val	Tyr	Ile	Val	Phe	Leu	Ala	Glu	Asn	Val	125	130	135
Lys	Gln	Val	His	Glu	Gly	Phe	Leu	Glu	Ser	Lys	Val	Phe	Ile	Ser	140	145	150
Asn	Ser	Thr	Asn	Ser	Ser	Asn	Pro	Cys	Glu	Arg	Arg	Ser	Val	Asp	155	160	165
Leu	Arg	Ile	Tyr	Met	Leu	Cys	Phe	Leu	Pro	Phe	Ile	Ile	Leu	Leu	170	175	180
Val	Phe	Ile	Arg	Glu	Leu	Lys	Asn	Leu	Phe	Val	Leu	Ser	Phe	Leu	185	190	195
Ala	Asn	Val	Ser	Met	Ala	Val	Ser	Leu	Val	Ile	Ile	Tyr	Gln	Tyr	200	205	210
Val	Val	Arg	Asn	Met	Pro	Asp	Pro	His	Asn	Leu	Pro	Ile	Val	Ala	215	220	225
Gly	Trp	Lys	Lys	Tyr	Pro	Leu	Phe	Phe	Gly	Thr	Ala	Val	Phe	Ala	230	235	240
Phe	Glu	Gly	Ile	Gly	Val	Val	Leu	Pro	Leu	Glu	Asn	Gln	Met	Lys	245	250	255
Glu	Ser	Lys	Arg	Phe	Pro	Gln	Ala	Leu	Asn	Ile	Gly	Met	Gly	Ile	260	265	270
Val	Thr	Thr	Leu	Tyr	Val	Thr	Leu	Ala	Thr	Leu	Gly	Tyr	Met	Cys	275	280	285
Phe	His	Asp	Glu	Ile	Lys	Gly	Ser	Ile	Thr	Leu	Asn	Leu	Pro	Gln	290	295	300
Asp	Val	Trp	Leu	Tyr	Gln	Ser	Val	Lys	Ile	Leu	Tyr	Ser	Phe	Gly	305	310	315
Ile	Phe	Val	Thr	Tyr	Ser	Ile	Gln	Phe	Tyr	Val	Pro	Ala	Glu	Ile	320	325	330
Ile	Ile	Pro	Gly	Ile	Thr	Ser	Lys	Phe	His	Thr	Lys	Trp	Lys	Gln	335	340	345
Ile	Cys	Glu	Phe	Gly	Ile	Arg	Ser	Phe	Leu	Val	Ser	Ile	Thr	Cys	350	355	360
Ala	Gly	Ala	Ile	Leu	Ile	Pro	Arg	Leu	Asp	Ile	Val	Ile	Ser	Phe	365	370	375
Val	Gly	Ala	Val	Ser	Ser	Ser	Thr	Leu	Ala	Leu	Ile	Leu	Pro	Pro	380	385	390
Leu	Val	Glu	Ile	Leu	Thr	Phe	Ser	Lys	Glu	His	Tyr	Asn	Ile	Trp	395	400	405
Met	Val	Leu	Lys	Asn	Ile	Ser	Ile	Ala	Phe	Thr	Gly	Val	Val	Gly	410	415	420
Phe	Leu	Leu	Gly	Thr	Tyr	Ile	Thr	Val	Glu	Glu	Ile	Ile	Tyr	Pro	425	430	435
Thr	Pro	Lys	Val	Val	Ala	Gly	Thr	Pro	Gln	Ser	Pro	Phe	Leu	Asn	440	445	450
Leu	Asn	Ser	Thr	Cys	Leu	Thr	Ser	Gly	Leu	Lys					455	460	465
															470	475	

<210> 24

<211> 237

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 2720354CD1

<400> 24

Met	Gly	Leu	Thr	Phe	Ile	Asn	Ala	Leu	Val	Phe	Gly	Val	Gln	Gly	1	5	10	15
Asn	Thr	Leu	Arg	Ala	Leu	Gly	His	Asp	Ser	Pro	Leu	Asn	Gln	Phe	20	25	30	
Leu	Ala	Gly	Ala	Ala	Ala	Gly	Ala	Ile	Gln	Cys	Val	Ile	Cys	Cys				

Pro Met Glu Leu	Ala Lys Thr Arg Leu	Gln Leu Gln Asp Ala	Gly	35	40	45
				50	55	60
Pro Ala Arg Thr	Tyr Lys Gly Ser Leu	Asp Cys Leu Ala Gln	Ile	65	70	75
				80	85	90
Tyr Gly His Glu	Gly Leu Arg Gly Val	Asn Arg Gly Met Val	Ser	95	100	105
				110	115	120
Thr Leu Leu Arg	Glu Thr Pro Ser Phe	Gly Val Tyr Phe Leu	Thr	125	130	135
				140	145	150
Tyr Asp Ala Leu	Thr Arg Ala Leu Gly	Cys Glu Pro Gly Asp	Arg	155	160	165
				170	175	180
Leu Leu Val Pro	Lys Leu Leu Leu Ala	Gly Gly Thr Ser Gly	Ile	185	190	195
				200	205	210
Val Ser Trp Leu	Ser Thr Tyr Pro Val	Asp Val Val Lys Ser	Arg	215	220	225
				230	235	
Leu Gln Ala Asp	Gly Leu Arg Gly Ala	Pro Arg Tyr Arg Gly	Ile			
Leu Asp Cys Val	His Gln Ser Tyr Arg	Ala Glu Gly Trp Arg	Val			
Phe Thr Arg Gly	Leu Ala Ser Thr Leu	Leu Arg Ala Phe Pro	Val			
Asn Ala Ala Thr	Phe Ala Thr Val Thr	Val Val Leu Thr Tyr	Ala			
Arg Gly Glu Glu	Ala Gly Pro Glu Gly	Glu Ala Val Pro Ala	Ala			
Pro Ala Gly Pro	Ala Leu Ala Gln Pro	Ser Ser Leu				

<210> 25

<211> 345

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 3038193CD1

<400> 25

Met Arg Leu Leu	Glu Arg Met Arg Lys	Asp Trp Phe Met Val	Gly	1	5	10	15
				20	25	30	35
Ile Val Leu Ala	Ile Ala Gly Ala Lys	Leu Glu Pro Ser Ile	Gly	40	45	50	55
				60	65	70	75
Val Asn Gly Gly	Pro Leu Lys Pro Glu	Ile Thr Val Ser Tyr	Ile	80	85	90	95
				100	105	110	115
Ala Val Ala Thr	Ile Phe Phe Asn Ser	Gly Leu Ser Leu Lys	Thr	120	125	130	135
				140	145	150	155
Glu Glu Leu Thr	Ser Ala Leu Val His	Leu Lys Leu His Leu	Phe	160	165	170	175
				180	185	190	195
Ile Gln Ile Phe	Thr Leu Ala Phe Phe	Pro Ala Thr Ile Trp	Leu	200	205	210	215
				220	225	230	235
Phe Leu Gln Leu	Leu Ser Ile Thr Pro	Ile Asn Glu Trp Leu	Leu	240	245	250	255
				260	265	270	275
Lys Gly Leu Gln	Thr Val Gly Cys Met	Pro Pro Pro Val Ser	Ser	280	285	290	295
				300	305	310	315
Ala Val Ile Leu	Thr Lys Ala Val Gly	Gly Asn Glu Gly Ile	Val	320	325	330	335
				340	345	350	355
Ile Thr Pro Leu	Leu Leu Leu Phe Leu	Gly Ser Ser Ser Ser	Ser	360	365	370	375
				380	385	390	395
Val Pro Phe Thr	Ser Ile Phe Ser Gln	Leu Phe Met Thr Val	Val	400	405	410	415
				420	425	430	435
Val Pro Leu Ile	Ile Gly Gln Ile Val	Arg Arg Tyr Ile Lys	Asp	440	445	450	455
				460	465	470	475
Trp Leu Glu Arg	Lys Lys Pro Pro Phe	Gly Ala Ile Ser Ser	Ser	480	485	490	495
				500	505	510	515
Val Leu Leu Met	Ile Ile Tyr Thr Thr	Phe Cys Asp Thr Phe	Ser	520	525	530	535
				540	545	550	555
Asn Pro Asn Ile	Asp Leu Asp Lys Phe	Ser Leu Val Leu Ile	Leu	560	565	570	575

Phe Ile Ile Phe	215	Ser Ile Gln Leu Ser	220	Phe Met Leu Leu Thr	225
	230		235		240
Ile Phe Ser Thr	245	Arg Asn Asn Ser Gly	250	Phe Thr Pro Ala Asp	255
Val Ala Ile Ile	260	Phe Cys Ser Thr His	265	Ser Leu Thr Leu Gly	270
Ile Pro Met Leu	275	Lys Ile Val Phe Ala Gly	280	Tyr Glu His Leu Ser	285
Leu Ile Ser Val	290	Pro Leu Leu Ile Tyr	295	His Pro Ala Gln Ile Leu	300
Leu Gly Ser Val	305	Leu Val Pro Thr Ile	310	Lys Ser Trp Met Val Ser	315
Arg Gln Lys Lys	320	Leu Leu Gln Thr Arg	325	Gly Pro Leu Ala Asn Leu	330
Asn Asn Pro Glu	335	Gly Leu Glu Tyr Leu	340	Ser Ile Lys Phe Gly His	345

<210> 26

<211> 521

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 3460979CD1

<400> 26

Met Ala Ala Leu Ala	1	Pro Val Gly Ser	10	Pro Ala Ser Arg Gly	15
Arg Leu Ala Ala Gly	20	Leu Arg Leu Leu	25	Pro Met Leu Gly Leu	30
Gln Leu Leu Ala Glu	35	Pro Gly Leu Gly	40	Arg Val His His Leu	45
Leu Lys Asp Asp Val	50	Arg His Lys Val	55	His Leu Asn Thr Phe	60
Phe Phe Lys Asp Gly	65	Tyr Met Val Val	70	Asn Val Ser Ser Leu	75
Leu Asn Glu Pro Glu	80	Asp Lys Asp Val	85	Thr Ile Gly Phe Ser	90
Asp Arg Thr Lys Asn	95	Asp Gly Phe Ser	100	Ser Tyr Leu Asp Glu	105
Val Asn Tyr Cys Ile	110	Leu Lys Lys Gln	115	Ser Val Ser Val Thr	120
Leu Ile Leu Asp Ile	125	Ser Arg Ser Glu	130	Val Arg Val Lys Ser	135
Pro Glu Ala Gly Thr	140	Gln Leu Pro Lys	145	Ile Ile Phe Ser Arg	150
Glu Lys Val Leu Gly	155	Gln Ser Gln Glu	160	Pro Asn Val Asn Pro	165
Ser Ala Gly Asn Gln	170	Thr Gln Lys Thr	175	Gln Asp Gly Gly Lys	180
Lys Arg Ser Thr Val	185	Asp Ser Lys Ala	190	Met Gly Glu Lys Ser	195
Ser Val His Asn Asn	200	Gly Gly Ala Val	205	Ser Phe Gln Phe Phe	210
Asn Ile Ser Thr Asp	215	Asp Gln Glu Gly	220	Leu Tyr Ser Leu Tyr	225
His Lys Cys Leu Gly	230	Lys Glu Leu Pro	235	Ser Asp Lys Phe Thr	240
Ser Leu Asp Ile Glu	245	Ile Thr Glu Lys	250	Asn Pro Asp Ser Tyr	255
Ser Ala Gly Glu Ile	260	Pro Leu Pro Lys	265	Leu Tyr Ile Ser Met	270
Phe Phe Phe Phe Leu	275	Ser Gly Thr Ile	280	Trp Ile His Ile Leu	285

Lys	Arg	Arg	Asn	Asp	Val	Phe	Lys	Ile	His	Trp	Leu	Met	Ala	Ala	
				290					295					300	
Leu	Pro	Phe	Thr	Lys	Ser	Leu	Ser	Leu	Val	Phe	His	Ala	Ile	Asp	
				305					310					315	
Tyr	His	Tyr	Ile	Ser	Ser	Gln	Gly	Phe	Pro	Ile	Glu	Gly	Trp	Ala	
				320					325					330	
Val	Val	Tyr	Tyr	Ile	Thr	His	Leu	Leu	Lys	Gly	Ala	Leu	Leu	Phe	
				335					340					345	
Ile	Thr	Ile	Ala	Leu	Ile	Gly	Thr	Gly	Trp	Ala	Phe	Ile	Lys	His	
				350					355					360	
Ile	Leu	Ser	Asp	Lys	Asp	Lys	Lys	Ile	Phe	Met	Ile	Val	Ile	Pro	
				365					370					375	
Leu	Gln	Val	Leu	Ala	Asn	Val	Ala	Tyr	Ile	Ile	Ile	Glu	Ser	Thr	
				380					385					390	
Glu	Glu	Gly	Thr	Thr	Glu	Tyr	Gly	Leu	Trp	Lys	Asp	Ser	Leu	Phe	
				395					400					405	
Leu	Val	Asp	Leu	Leu	Cys	Cys	Gly	Ala	Ile	Leu	Phe	Pro	Val	Val	
				410					415					420	
Trp	Ser	Ile	Arg	His	Leu	Gln	Glu	Ala	Ser	Ala	Thr	Asp	Gly	Lys	
				425					430					435	
Ala	Ala	Ile	Asn	Leu	Ala	Lys	Leu	Lys	Leu	Phe	Arg	His	Tyr	Tyr	
				440					445					450	
Val	Leu	Ile	Val	Cys	Tyr	Ile	Tyr	Phe	Thr	Arg	Ile	Ile	Ala	Phe	
				455					460					465	
Leu	Leu	Lys	Leu	Ala	Val	Pro	Phe	Gln	Trp	Lys	Trp	Leu	Tyr	Gln	
				470					475					480	
Leu	Leu	Asp	Glu	Thr	Ala	Thr	Leu	Val	Phe	Phe	Val	Leu	Thr	Gly	
				485					490					495	
Tyr	Lys	Phe	Arg	Pro	Ala	Ser	Asp	Asn	Pro	Tyr	Leu	Gln	Leu	Ser	
				500					505					510	
Gln	Glu	Glu	Glu	Asp	Leu	Glu	Met	Glu	Ser	Val					
				515					520						

<210> 27

<211> 555

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 7472200CD1

<400> 27

Met	Thr	Leu	Val	Tyr	Phe	Pro	Pro	Ser	Lys	Leu	Gln	Gln	Gln	Gln	
1				5					10					15	
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<213> Homo sapiens

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<223> Incyte ID No: 7472002CB1

<400> 34

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<212> DNA

<213> Homo sapiens

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<400> 37

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<210> 39
<211> 2043
<212> DNA
<213> Homo sapiens

<220>
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<223> Incyte ID No: 70064803CB1

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2043

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<210> 40

<211> 1915

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 70356768CB1

<400> 40

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<210> 41
<211> 1809
<212> DNA
<213> Homo sapiens

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<220>
<221> misc_feature
<223> Incyte ID No: 5674114CB1

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<210> 42
<211> 1730
<212> DNA
<213> Homo sapiens

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<220>
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<223> Incyte ID No: 1254635CB1

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<210> 43

<211> 1147

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1670595CB1

<400> 43

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<211> 2745

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1859560CB1

<400> 44

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